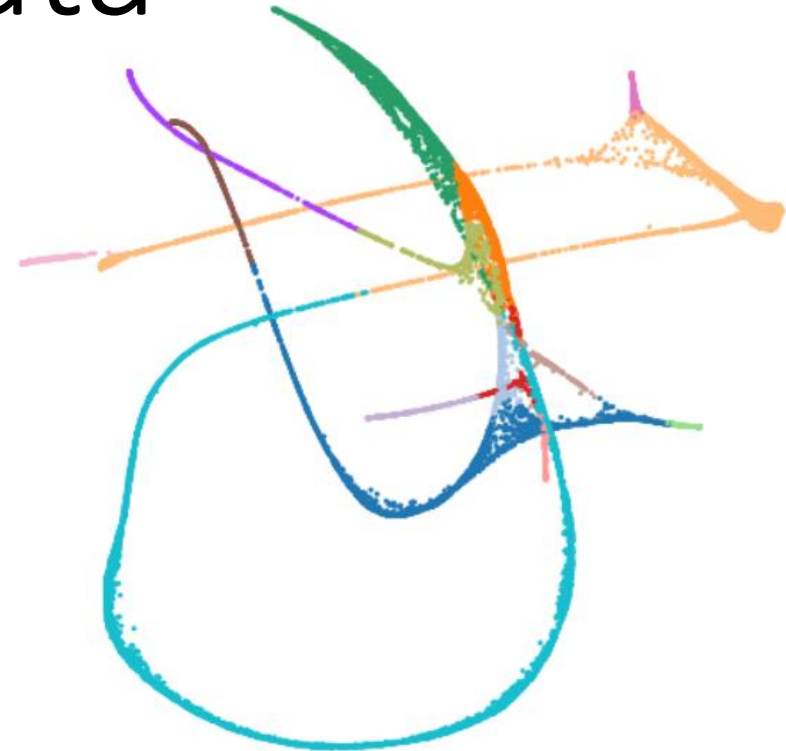
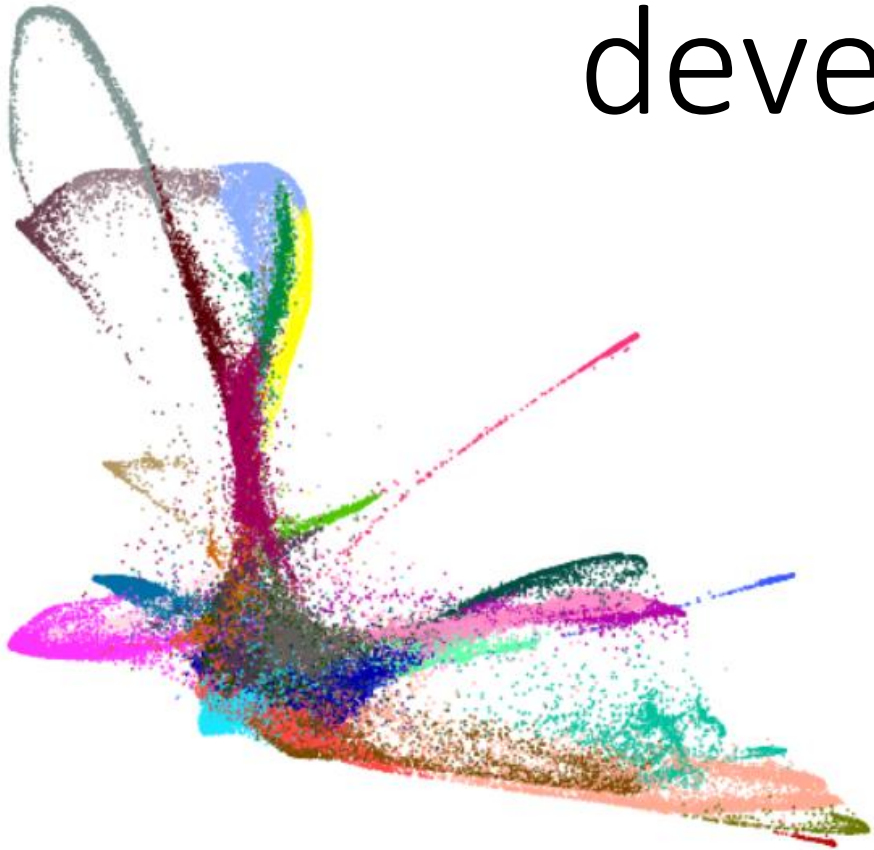


Deciphering dynamical developmental data

Louis Faure - PhD

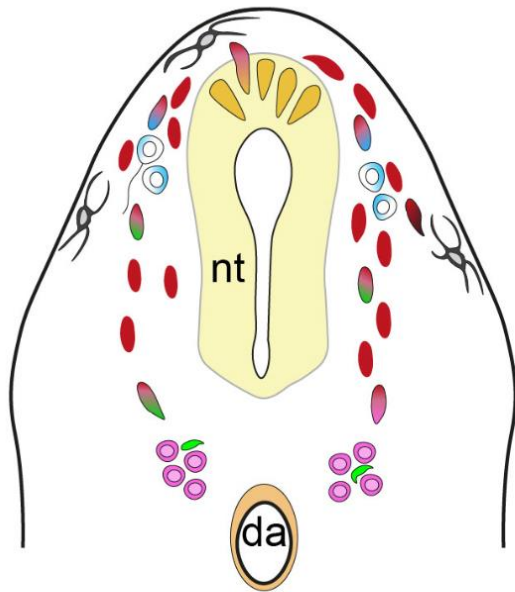
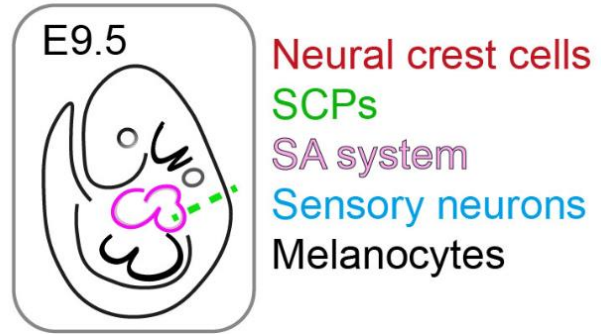


My background



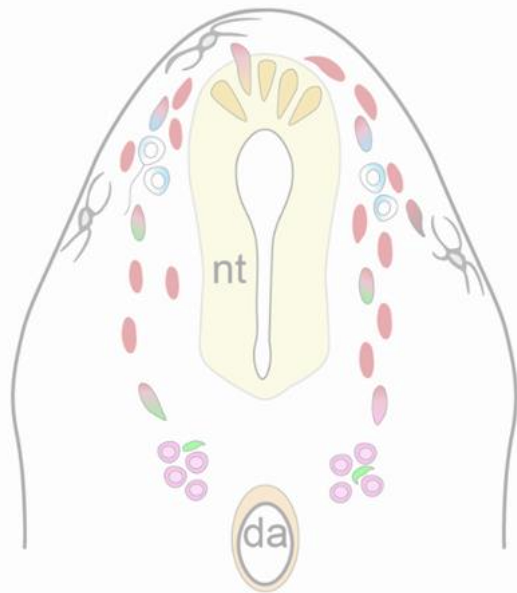
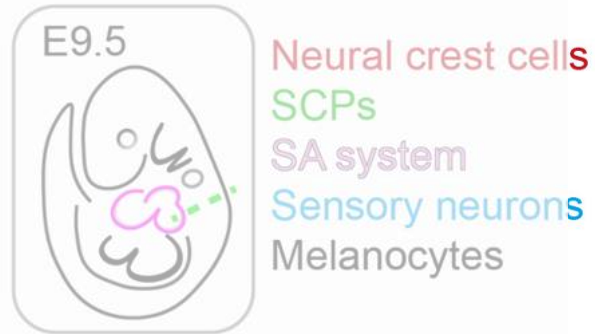
My PhD project

A Neural crest cell migration

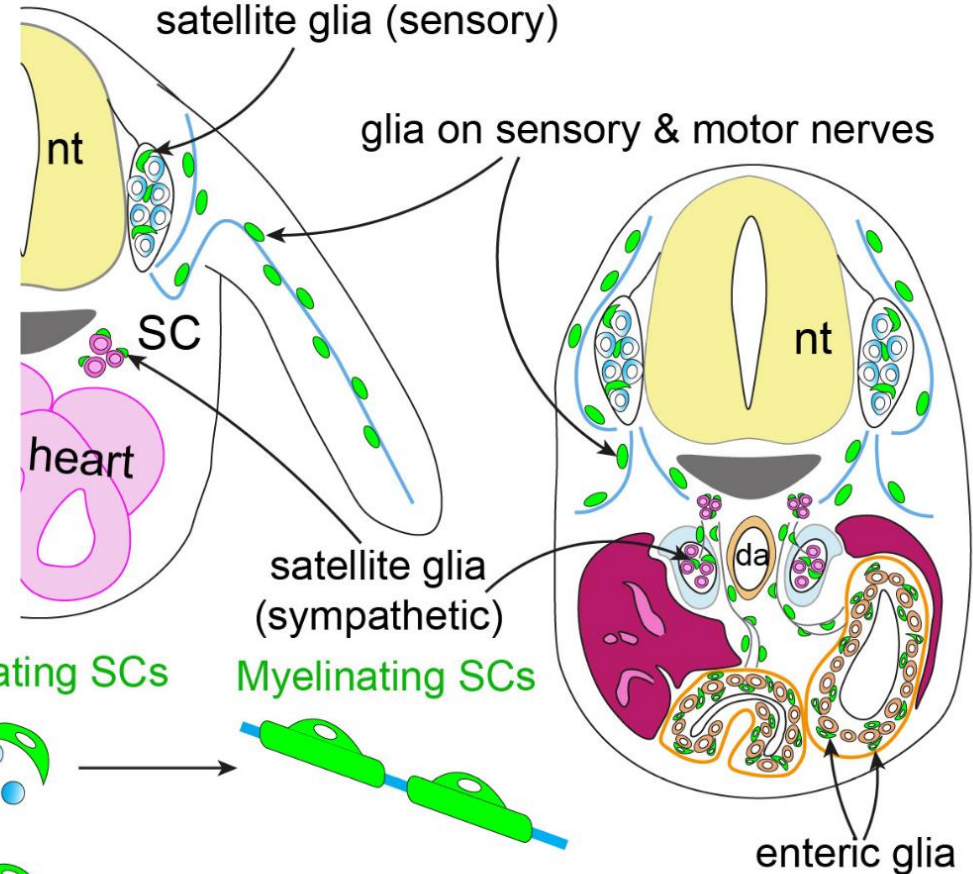
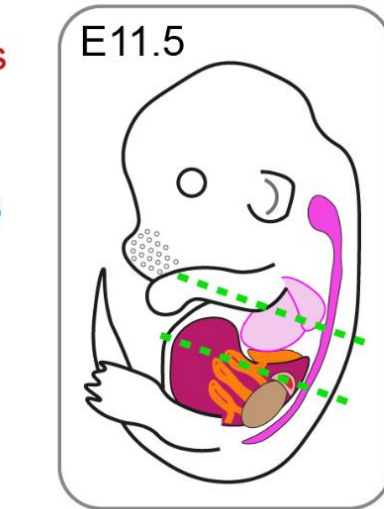


My PhD project

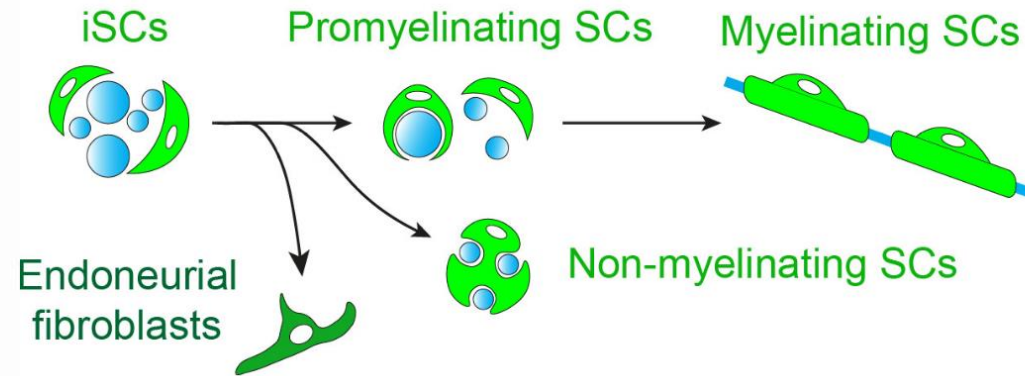
A Neural crest cell migration



Schwann cell precursor migration on peripheral nerves

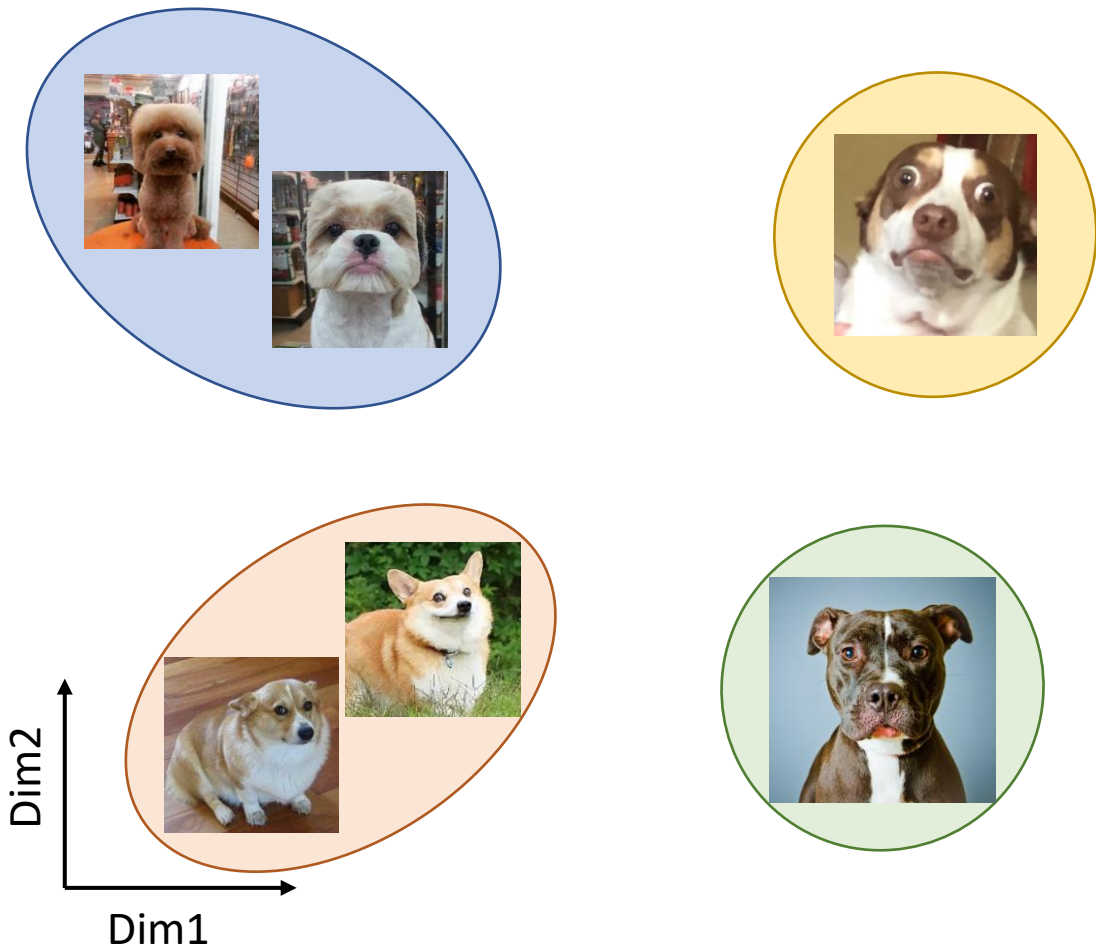


Myelinating lineage

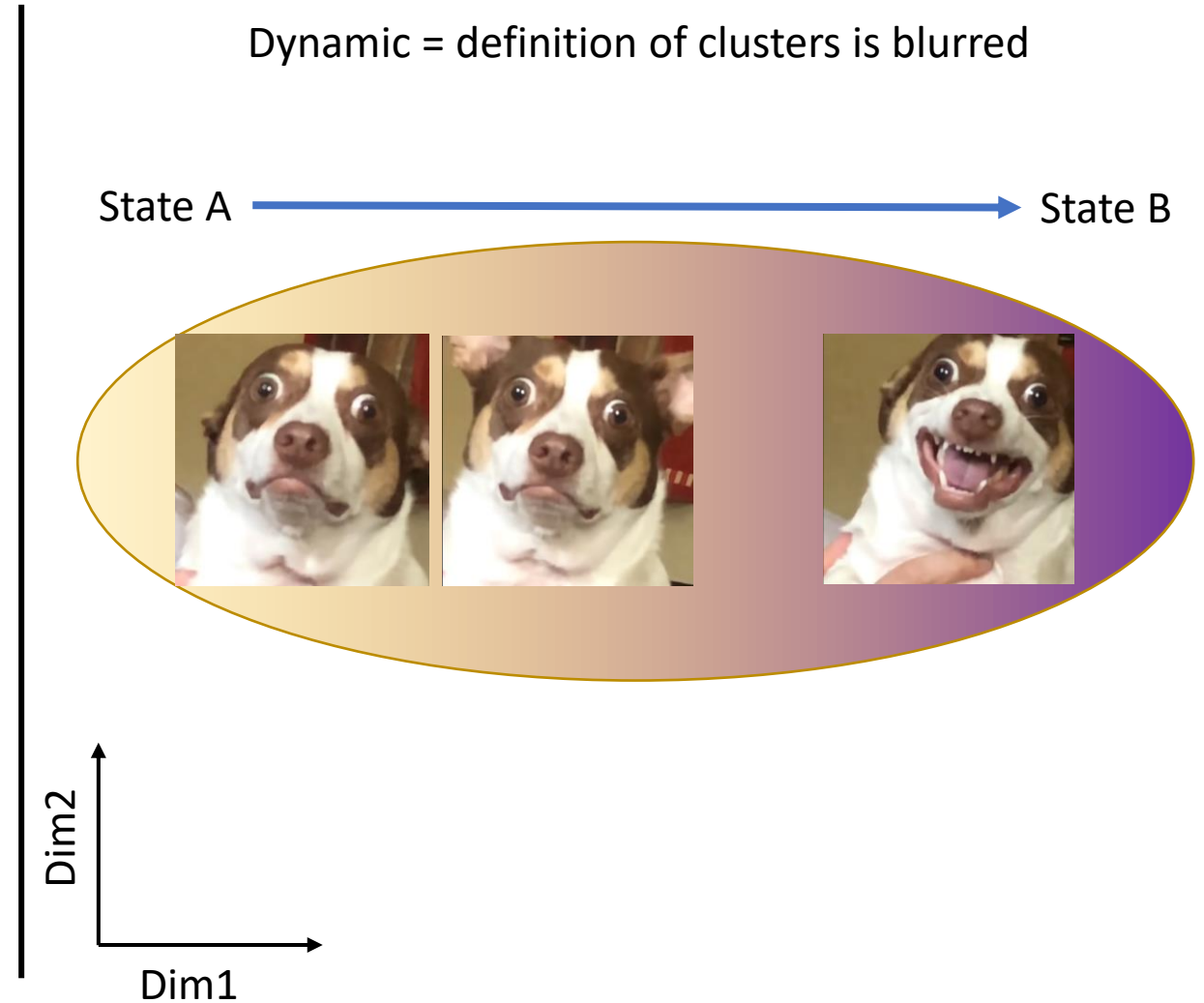


Static populations versus dynamical ones

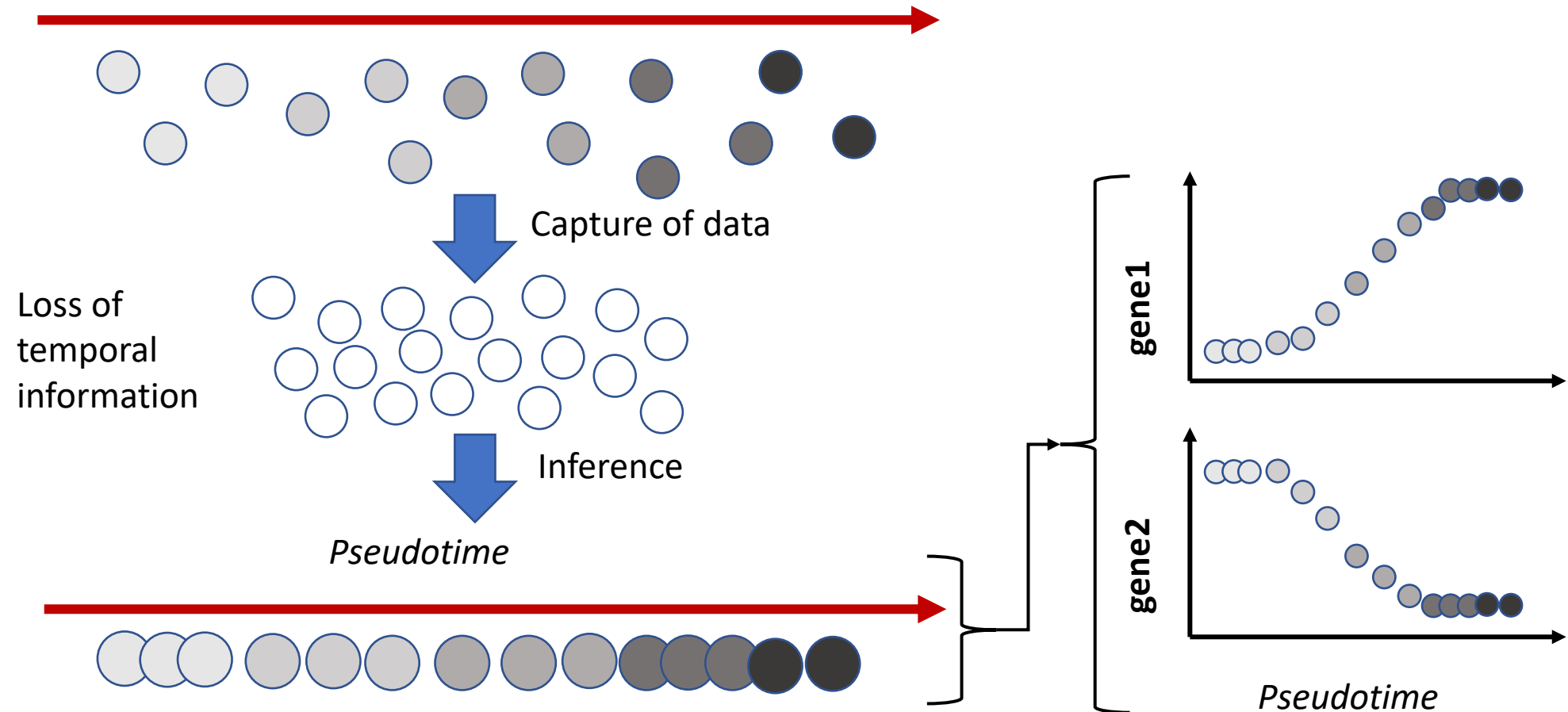
Static = well defined cell-type/clusters



Dynamic = definition of clusters is blurred



Concept of pseudotime ordering

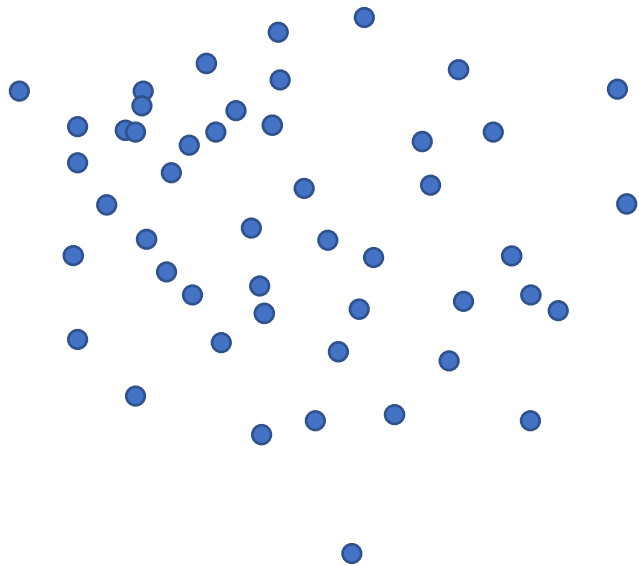
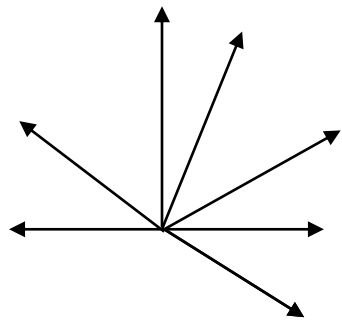


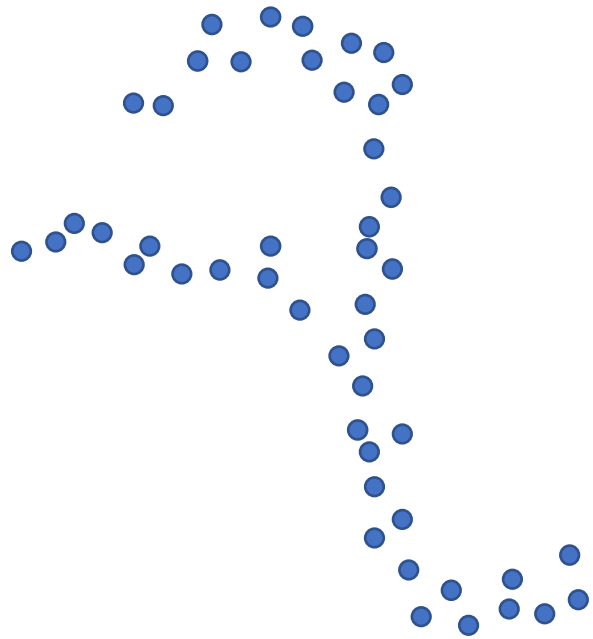
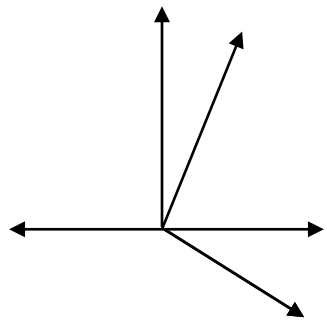
How do we apply this to scRNAseq data?

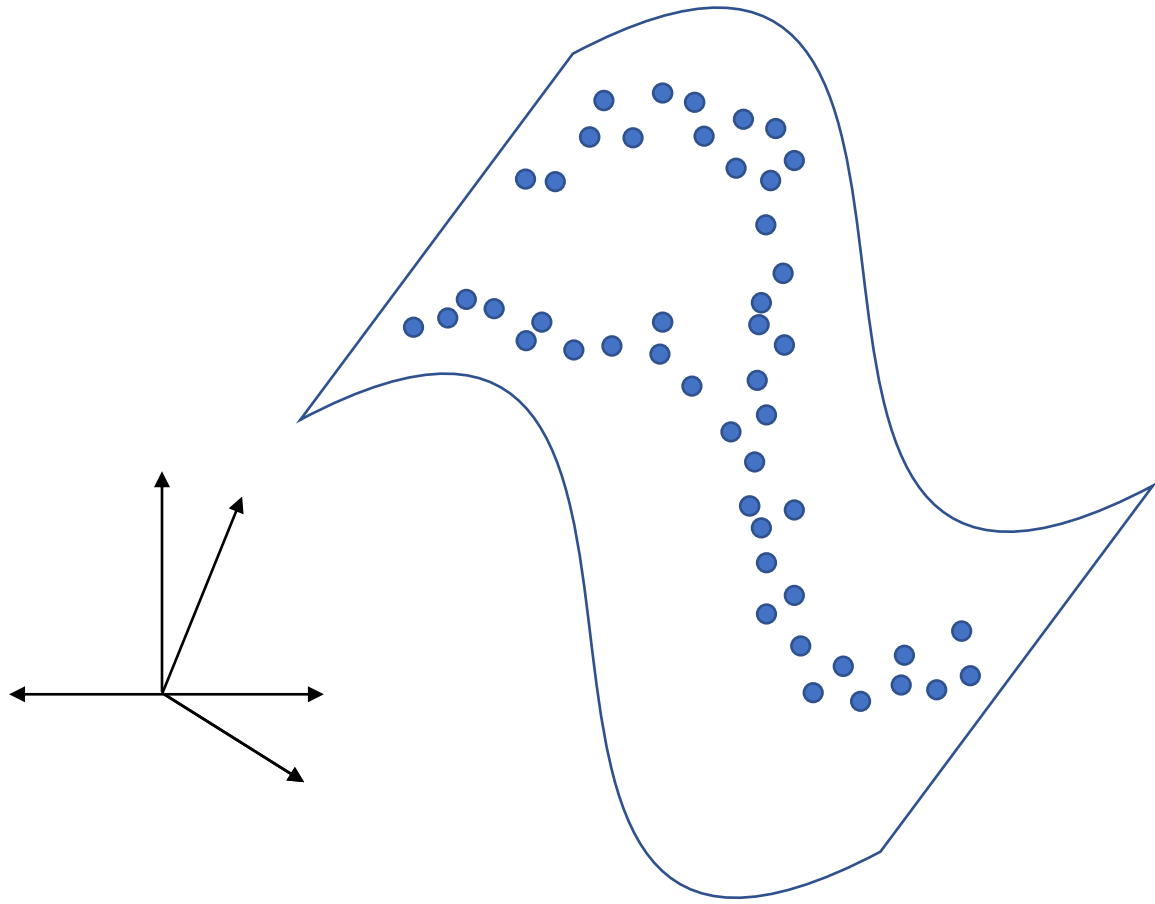
- scRNAseq is highly noisy data
 - Amplification bias
 - Library size differences
 - Sequencing depth
 - **Biology is stochastic and noisy by itself!**
- But the good news is that transcriptional activity happens in **modules composed numerous genes**
 - Latent structure exists at a much lower dimension

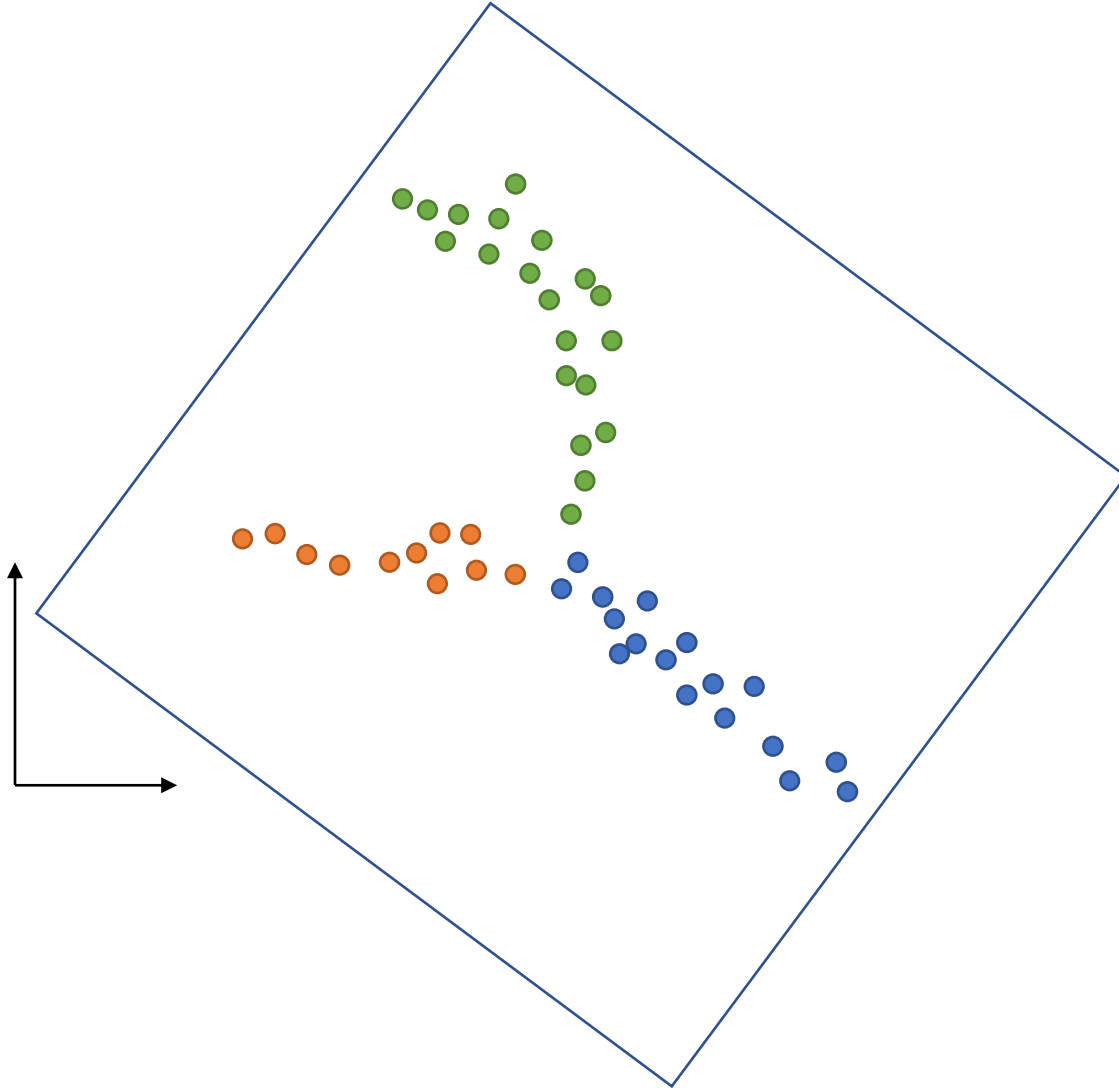
Additional processing steps are required to “denoise” and extract the latent structure of the data

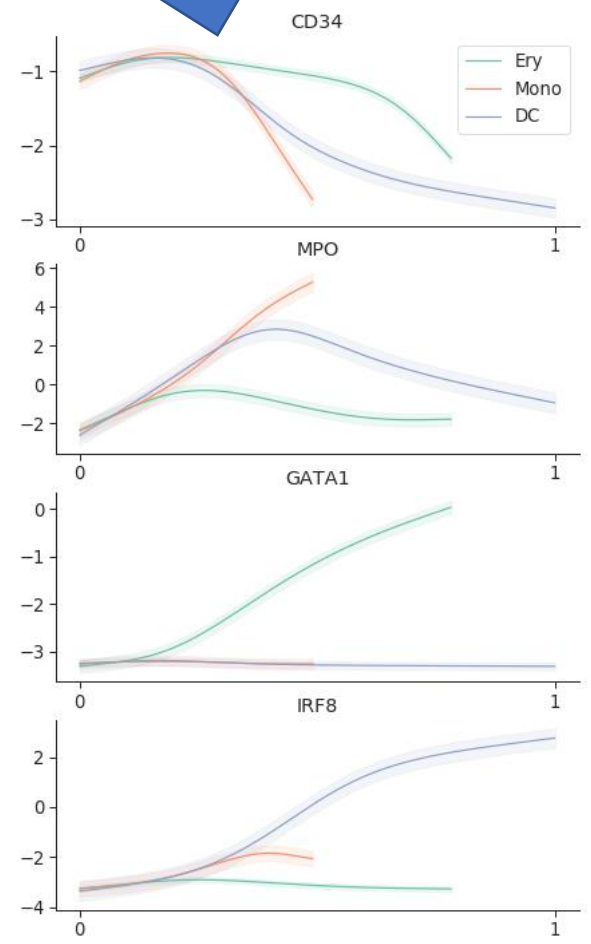
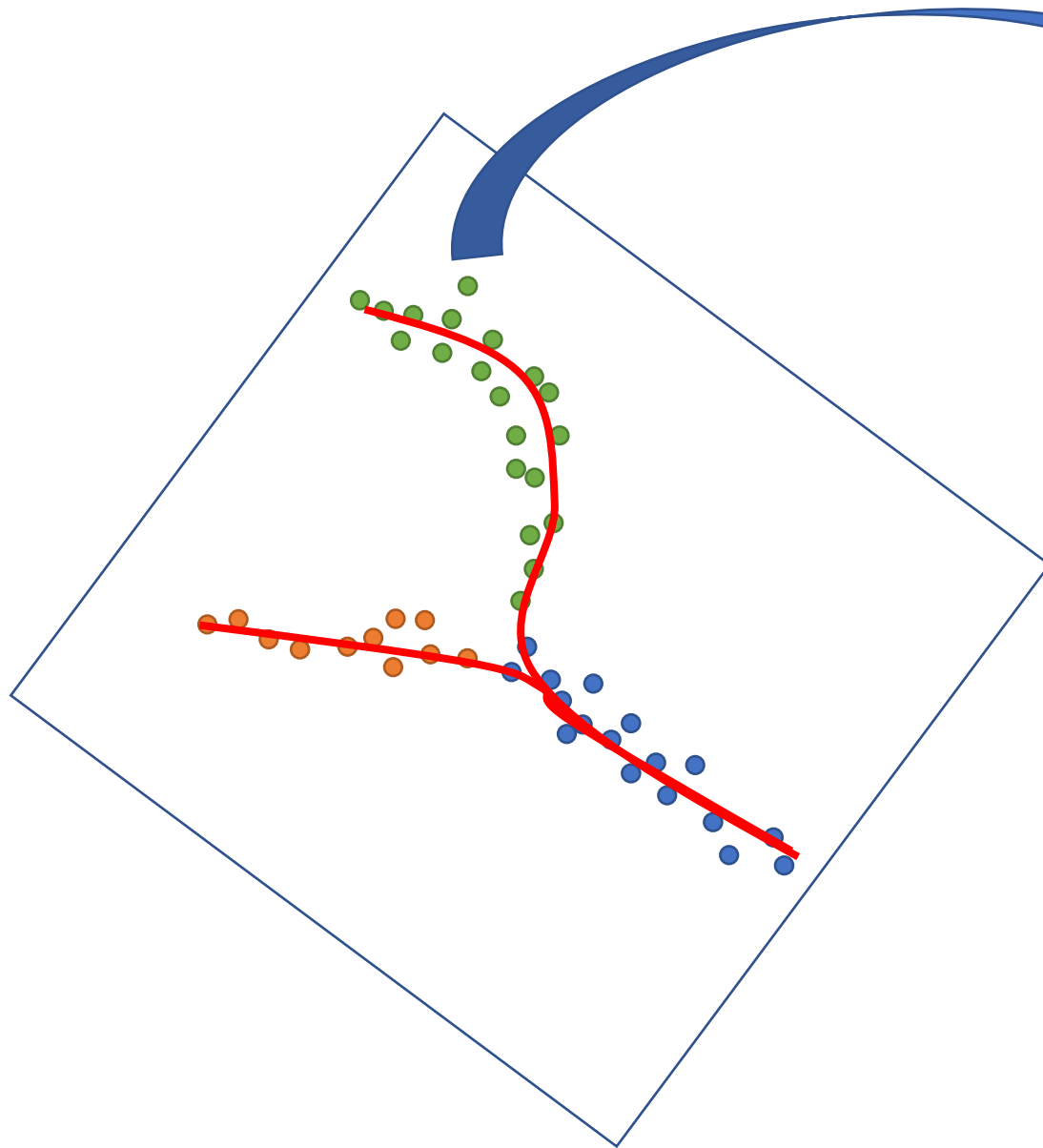












Denoising data: PCA

Highly dimensional data

n samples
 m features



Obtain **covariance matrix A**
of size $n \times n$



eigen decompose A
by diagonalizing:
 $A = XDX^{-1}$

D is a diagonal matrix
made of **eigenvalues**

$$\begin{bmatrix} \lambda_1 & 0 & 0 \\ 0 & \lambda_2 & 0 \\ 0 & 0 & \lambda_n \end{bmatrix}$$

Eigenvalues are found by
solving $|A - \lambda I| = 0$
where I is an identity matrix



X is a matrix with
each columns being
eigenvectors of A

Eigenvector \vec{x}_n are found by
solving $(A - \lambda I)\vec{x}_n = \vec{0}$ for
each eigenvalues λ_n previously
found.

Important!

- We have decomposed our data into linear transformations with:
 - the eigenvectors being the "axes" or the "directions"
 - the eigenvalues being the scaling factors
- Each eigenvector has its own eigenvalue
- By choosing a reduced number among the top eigenvalues, we subset our eigenvectors columns which becomes our **reduced dimensions!**

In this case, dimensions are principal components

Denoising data: PCA

“Highly” dimensional data

20 samples

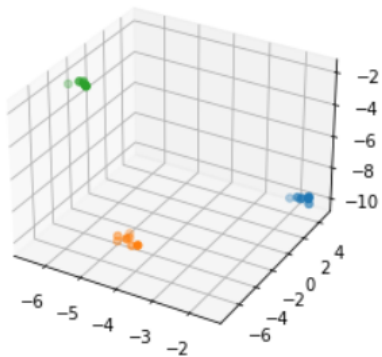
3 features

```
[1]: import numpy as np
      from sklearn.datasets import make_blobs
      blobs, clusters = make_blobs(n_samples=20,
                                  n_features=3,
                                  cluster_std=.2,
                                  random_state=1)
```

```
[2]: blobs = blobs[np.argsort(clusters),:]
      clusters = clusters[np.argsort(clusters)]
```

```
[3]: import matplotlib.pyplot as plt
      fig = plt.figure()
      ax = fig.add_subplot(111,
                           projection='3d')
```

```
      for c in range(3):
          ax.scatter(blobs[clusters==c,0],
                    blobs[clusters==c,1],
                    blobs[clusters==c,2])
```



https://github.com/LouisFaure/Trajectory_Inference_workshop/blob/main/01.%20Eigen-decomposition.ipynb

Obtain covariance matrix A of size $n \times n$ \longrightarrow eigen decompose A by diagonalizing:

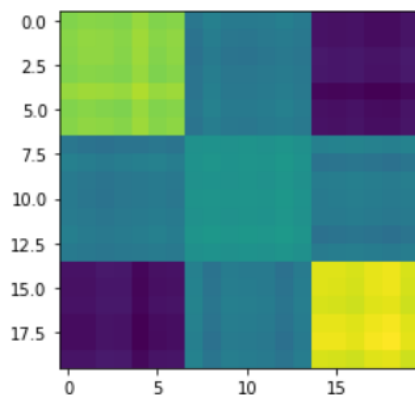
$$A = XDX^{-1}$$

**data is scaled to remove the effect of the magnitude of the variables*

```
[4]: from sklearn.preprocessing import *
      scaler = StandardScaler()
      blobs_scaled = scaler.fit_transform(blobs)
```

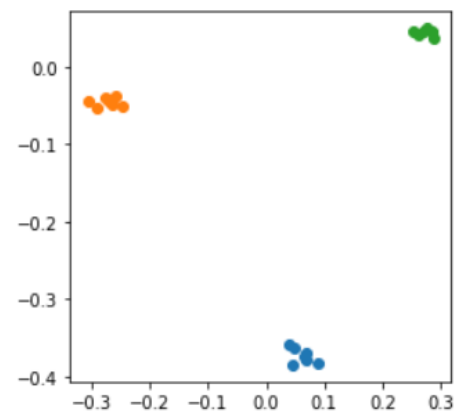
```
[5]: import numpy as np
      cov = np.cov(blobs_scaled)
```

```
[6]: plt.imshow(cov);
```



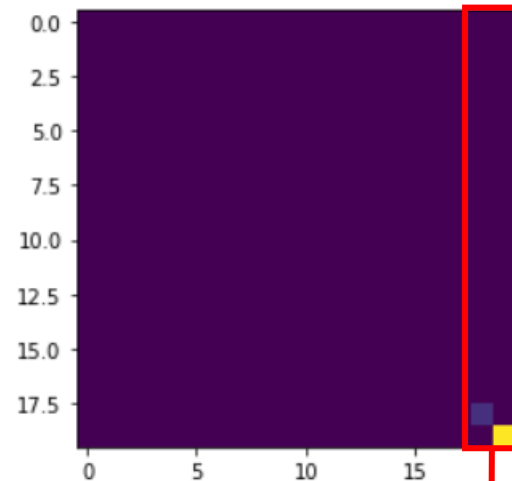
**the two last columns can be accessed via -2 and -1*

```
[9]: plt.figure(figsize=(4,4))
      for c in range(3):
          plt.scatter(X[clusters==c,-2],
                      X[clusters==c,-1])
```

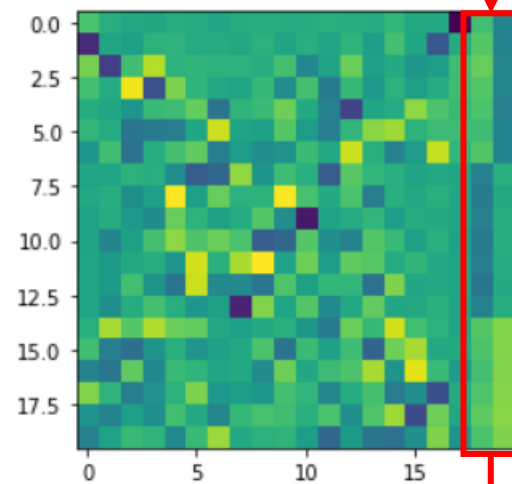


```
[7]: D, X = np.linalg.eigh(cov)
```

```
[8]: plt.imshow(np.diag(D));
```

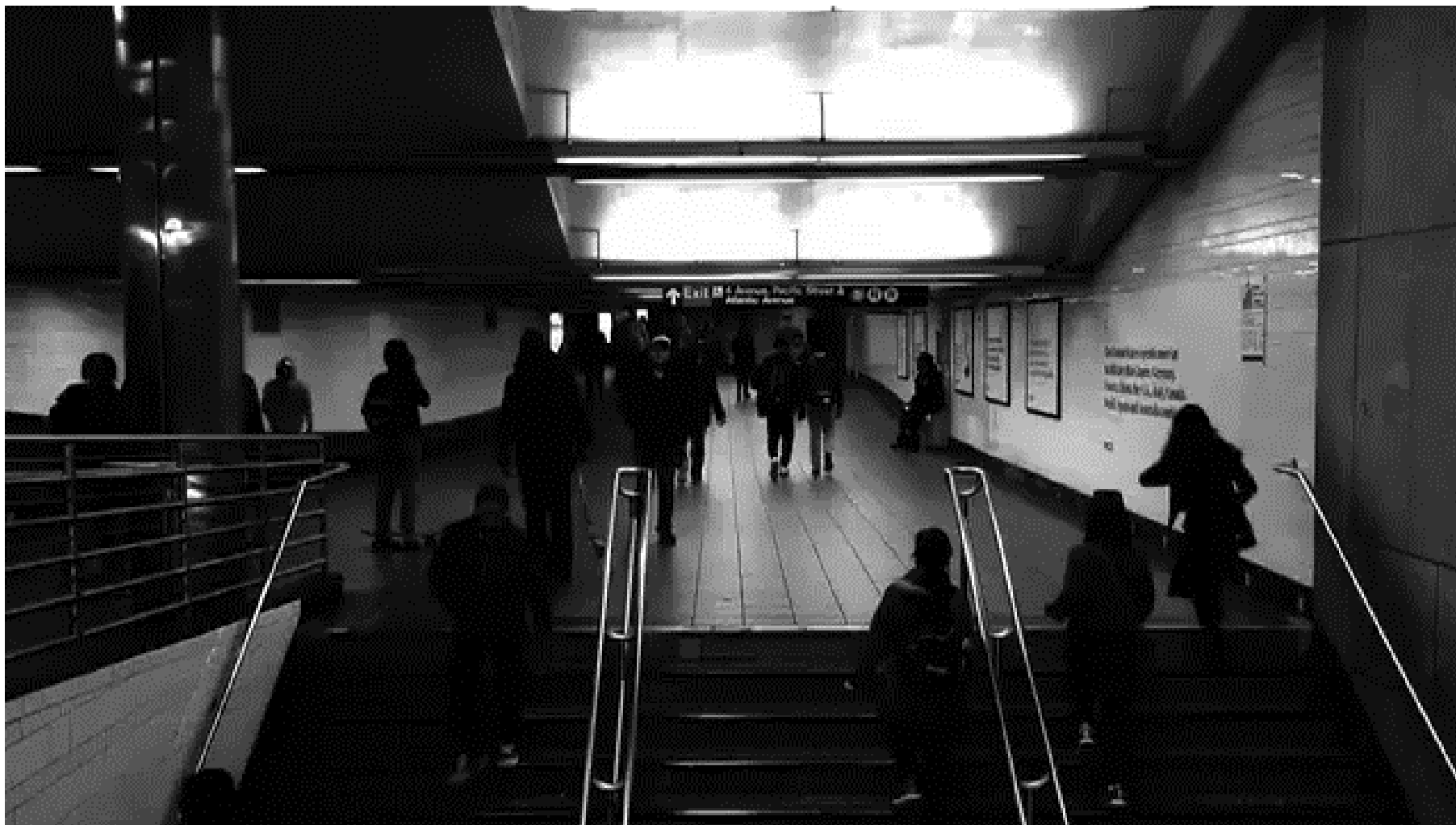


```
[9]: plt.imshow(X);
```

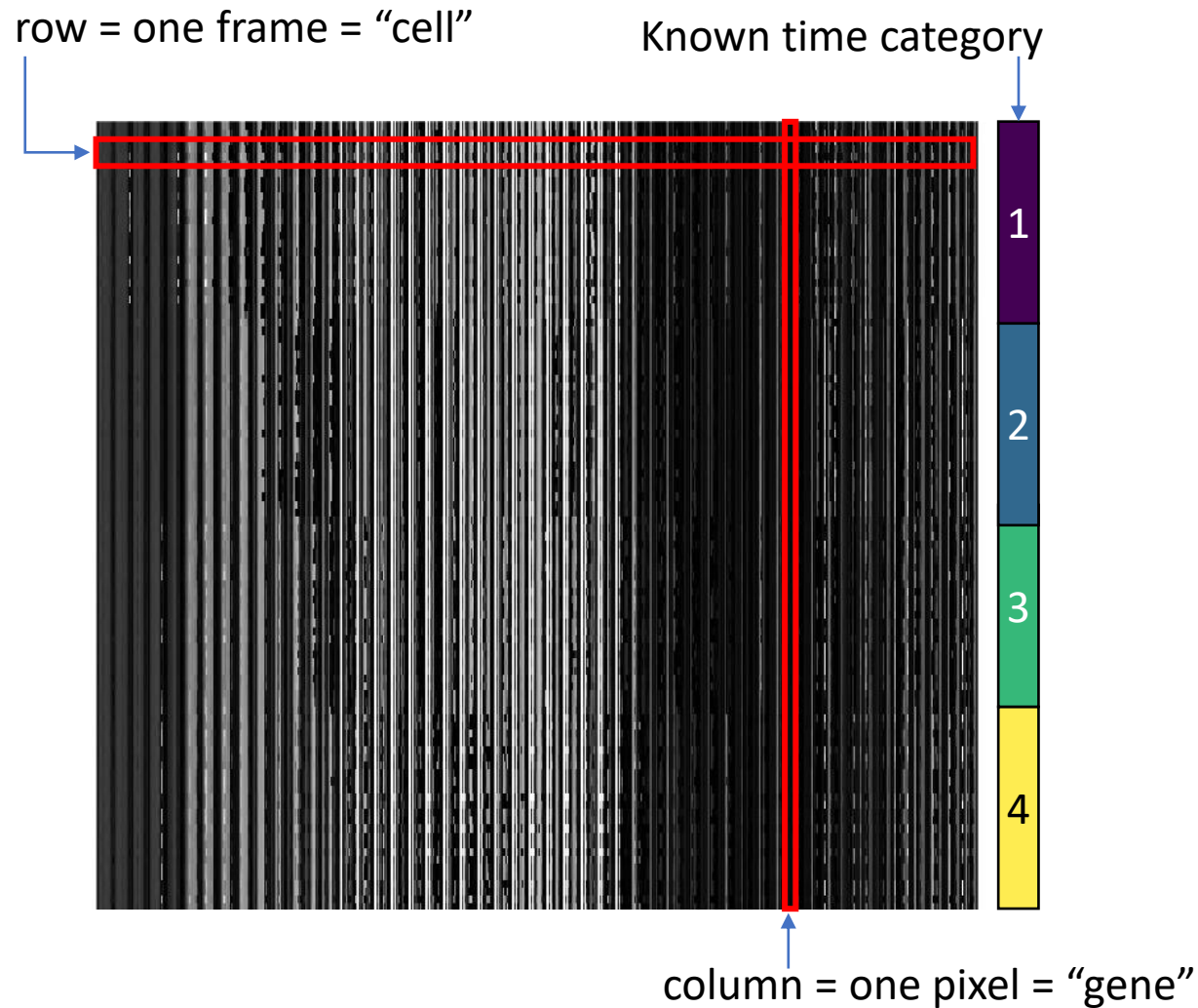


Select reduced dimensions

Pseudotime: recovering lost time information



Considering our video as our “count matrix”



```
[1]: import matplotlib.pyplot as plt
import matplotlib.image as mpimg
import numpy as np
import glob
import scanpy as sc

[2]: files=glob.glob('frames/*.jpg')

[3]: def rgb2gray(path):
    return np.dot(mpimg.imread(path)[...,:3], [0.2989, 0.5870, 0.1140]).ravel()

mat=np.vstack(list(map(rgb2gray, files)))

[4]: adata=sc.AnnData(mat)

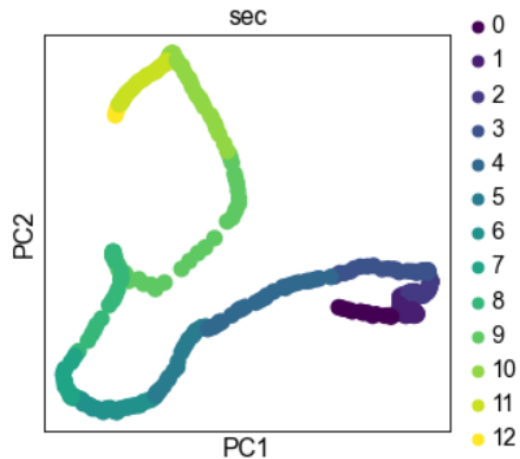
[5]: adata
AnnData object with n_obs × n_vars = 349 × 230400

[6]: adata.obs["sec"]=[round(int(f.split("_")[1].split(".")[0])/30) for f in files]
adata.obs["sec"]=adata.obs["sec"].astype("category")
```


Dimensionality reduction methods help revealing dynamical axis

Running PCA on scaled values

```
[7]: adata.obsm["X_pca"]=sc.pp.pca(sc.pp.scale(adata.X,copy=True),n_comps=20)
sc.set_figure_params()
sc.pl.pca(adata,color='sec',palette="viridis")
```



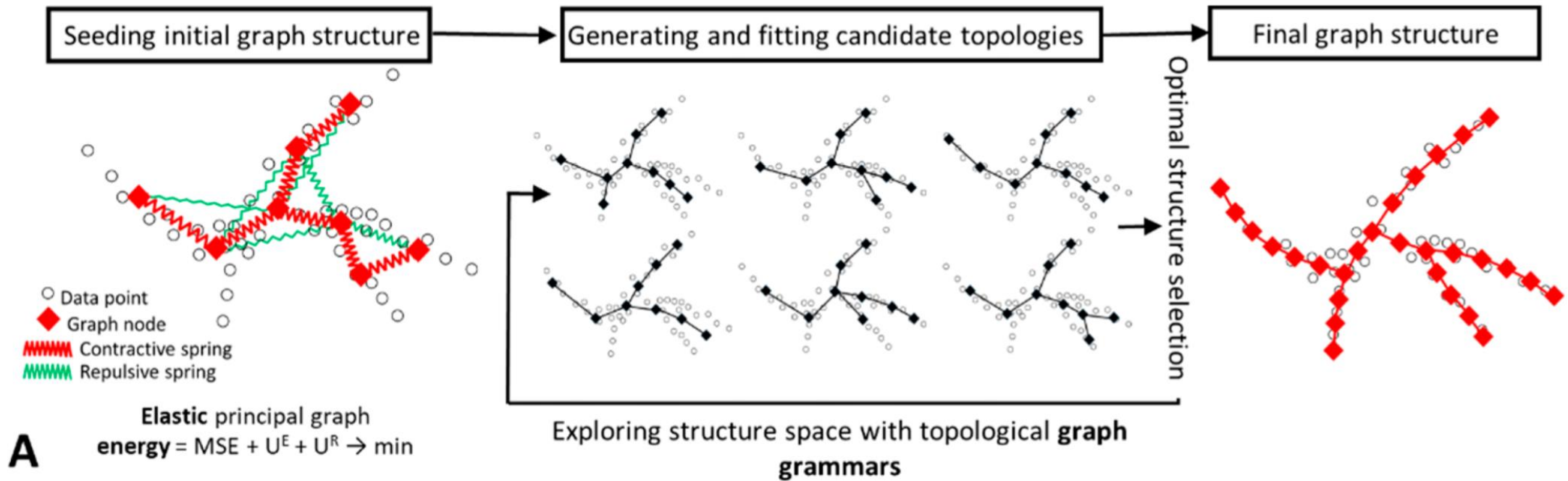
Infer a principal graph in PCA space

```
[9]: import scFates as scf
```

```
[10]: scf.tl.curve(adata,use_rep='X_pca',Nodes=30,seed=42)
```

```
inferring a principal tree --> parameters used
30 principal points, mu = 0.1, lambda = 0.01
finished (0:00:00) --> added
.uns['epg'] dictionary containing inferred elastic curve generated from elpigraph.
.uns['graph']['B'] adjacency matrix of the principal points.
.uns['graph']['R'] hard assignment of cells to principal point in representation space.
.uns['graph']['F'], coordinates of principal points in representation space.
```

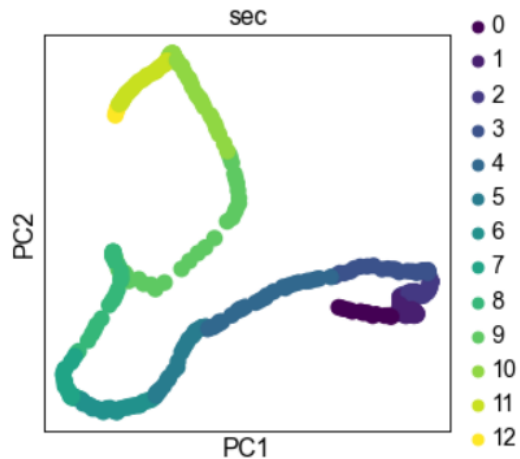
ELPiGraph



Dimensionality reduction methods help revealing dynamical axis

Running PCA on scaled values

```
[7]: adata.obsm["X_pca"]=sc.pp.pca(sc.pp.scale(adata.X,copy=True),n_comps=20)
sc.set_figure_params()
sc.pl.pca(adata,color='sec',palette="viridis")
```



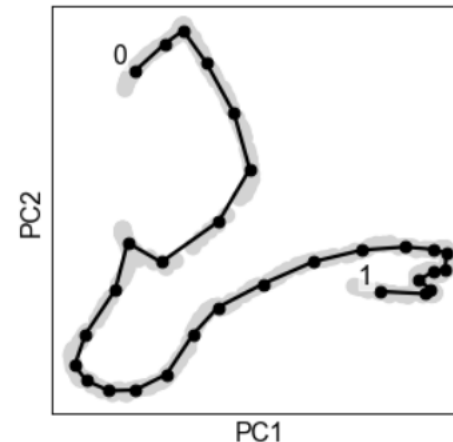
Infer a principal graph in PCA space

```
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```

```
[10]: scf.tl.curve(adata,use_rep='X_pca',Nodes=30,seed=42)
```

```
inferring a principal tree --> parameters used
30 principal points, mu = 0.1, lambda = 0.01
finished (0:00:00) --> added
.uns['epg'] dictionary containing inferred elastic curve generated from elpigraph.
.uns['graph']['B'] adjacency matrix of the principal points.
.uns['graph']['R'] hard assignment of cells to principal point in representation space.
.uns['graph']['F'], coordinates of principal points in representation space.
```

```
[11]: scf.pl.graph(adata,basis="pca")
```

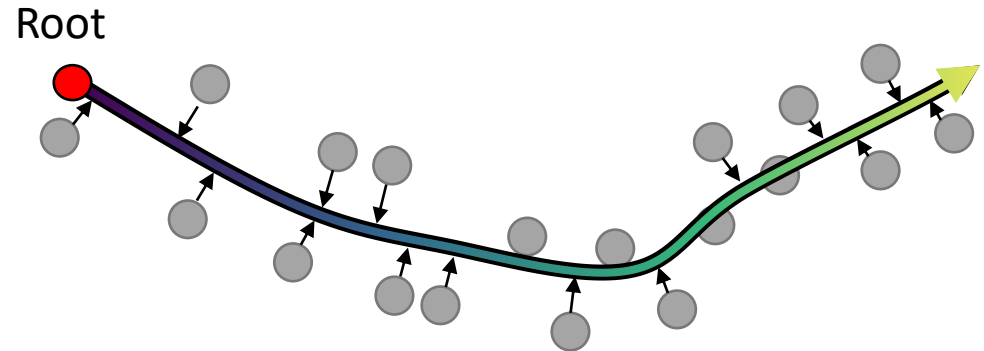
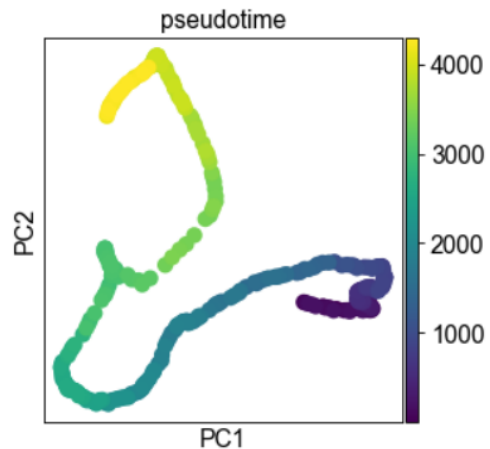


Pseudotime consists in reordering the cells along an inferred trajectory

```
[12]: scf.tl.root(adata,1)
root selected --> added
.uns['graph']['root'] selected root.
.uns['graph']['pp_info'] for each PP, its distance vs root and segment assignment.
.uns['graph']['pp_seg'] segments network information.
```

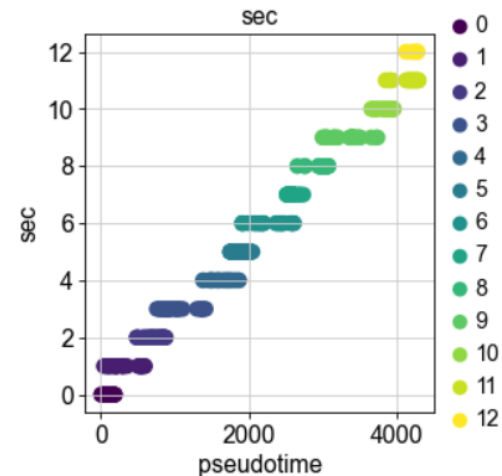
```
[13]: scf.tl.pseudotime(adata)
projecting cells onto the principal graph
finished (0:00:00) --> added
.obs['edge'] assigned edge.
.obs['t'] pseudotime value.
.obs['seg'] segment of the tree assigned.
.obs['milestones'] milestone assigned.
.uns['pseudotime_list'] list of cell projection from all mappings.
```

```
[14]: sc.pl.pca(adata,color="t",title='pseudotime')
```



```
[15]: ax=sc.pl.scatter(adata,x="t",y="sec",color="sec",show=False)
ax.set_xlabel("pseudotime")
```

```
[15]: Text(0.5, 0, 'pseudotime')
```

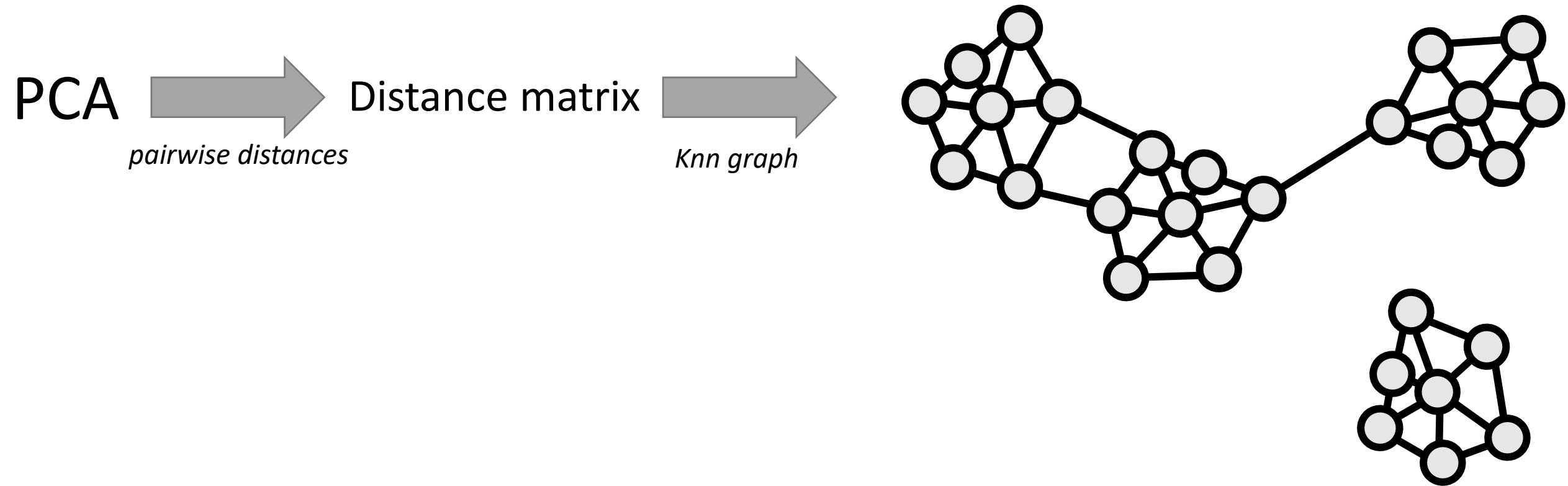


Recovered time! Yay!



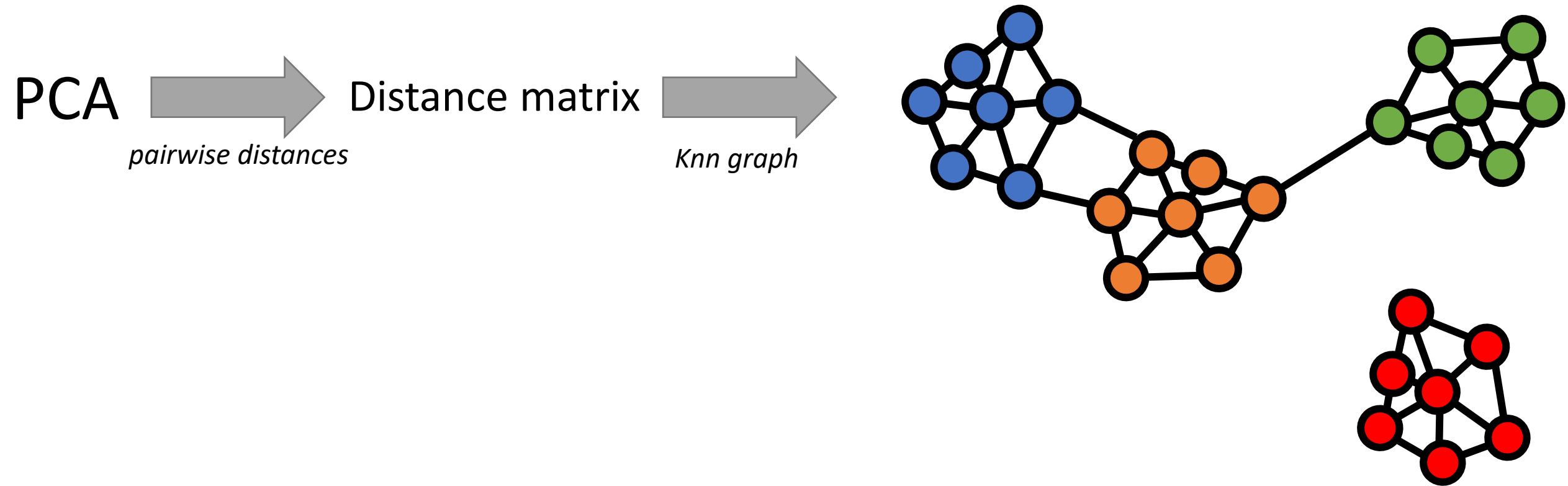
Denoising scRNAseq data: beyond PCA

Knn graph



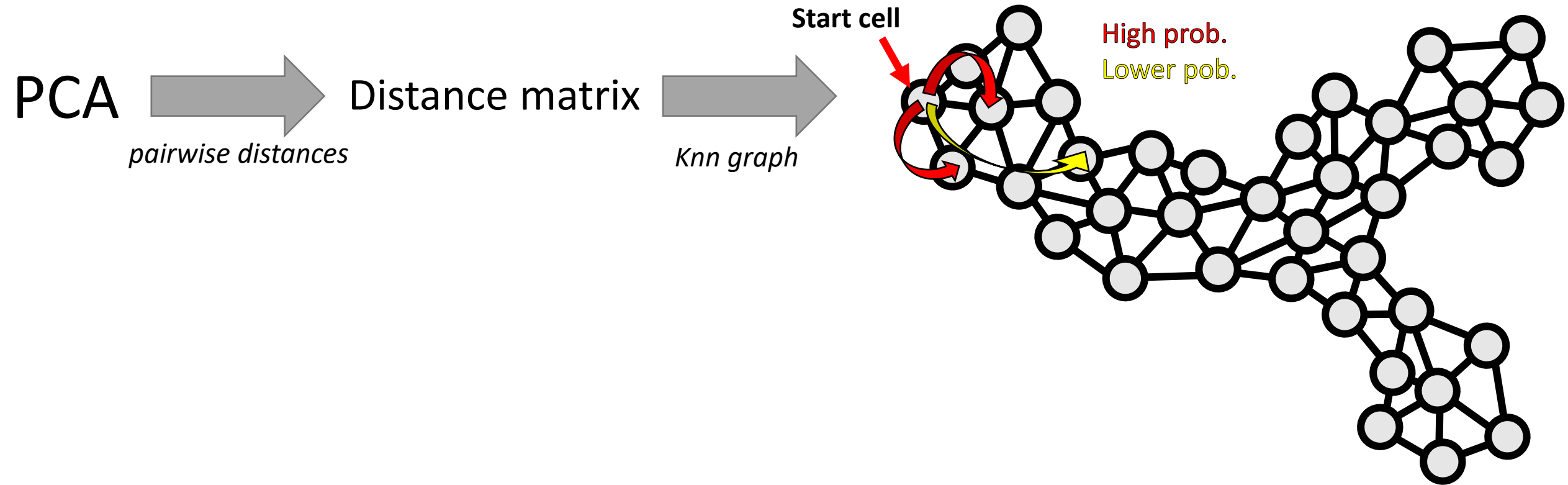
Denoising scRNAseq data: beyond PCA

Knn graph + clustering



Denoising developmental scRNAseq data

Knn graph

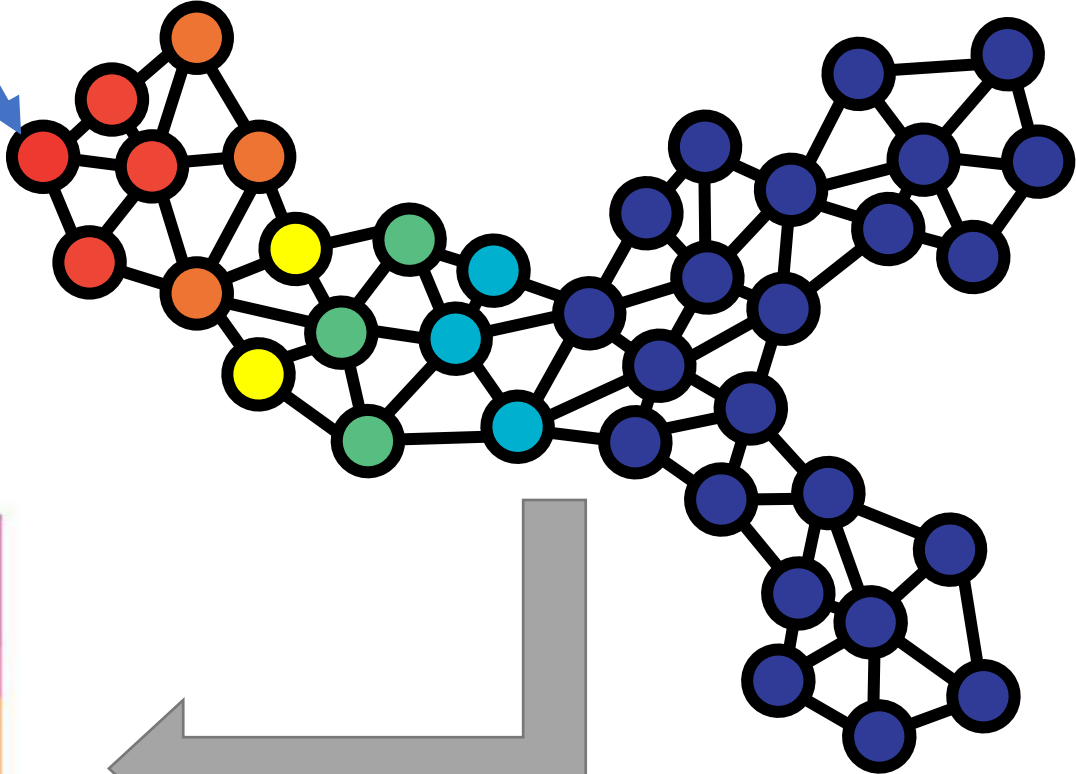


Denoising developmental scRNAseq data

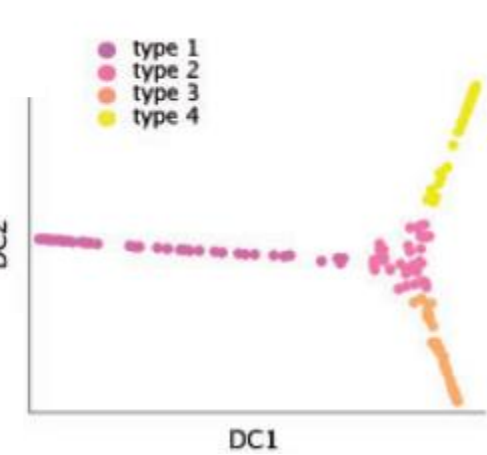
Knn graph

PCA $\xrightarrow{\text{pairwise distances}}$ Distance matrix

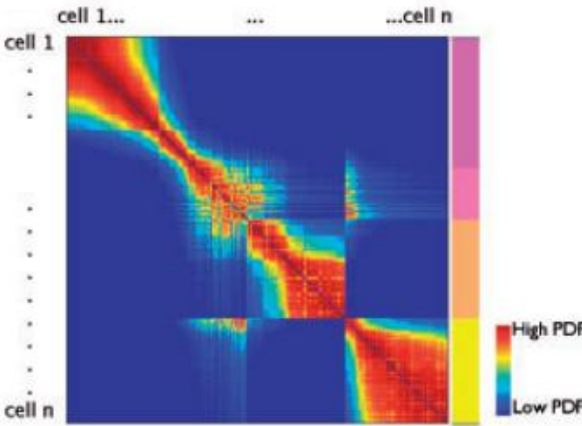
Distance matrix $\xrightarrow{\text{Knn graph}}$



Haghverdi, L., Buettner, F. & Theis, F. J. Diffusion maps for high-dimensional single-cell analysis of differentiation data. *Bioinformatics* (2015) doi:10.1093/bioinformatics/btv325.



$\xleftarrow{\text{Eigen decomposition}}$

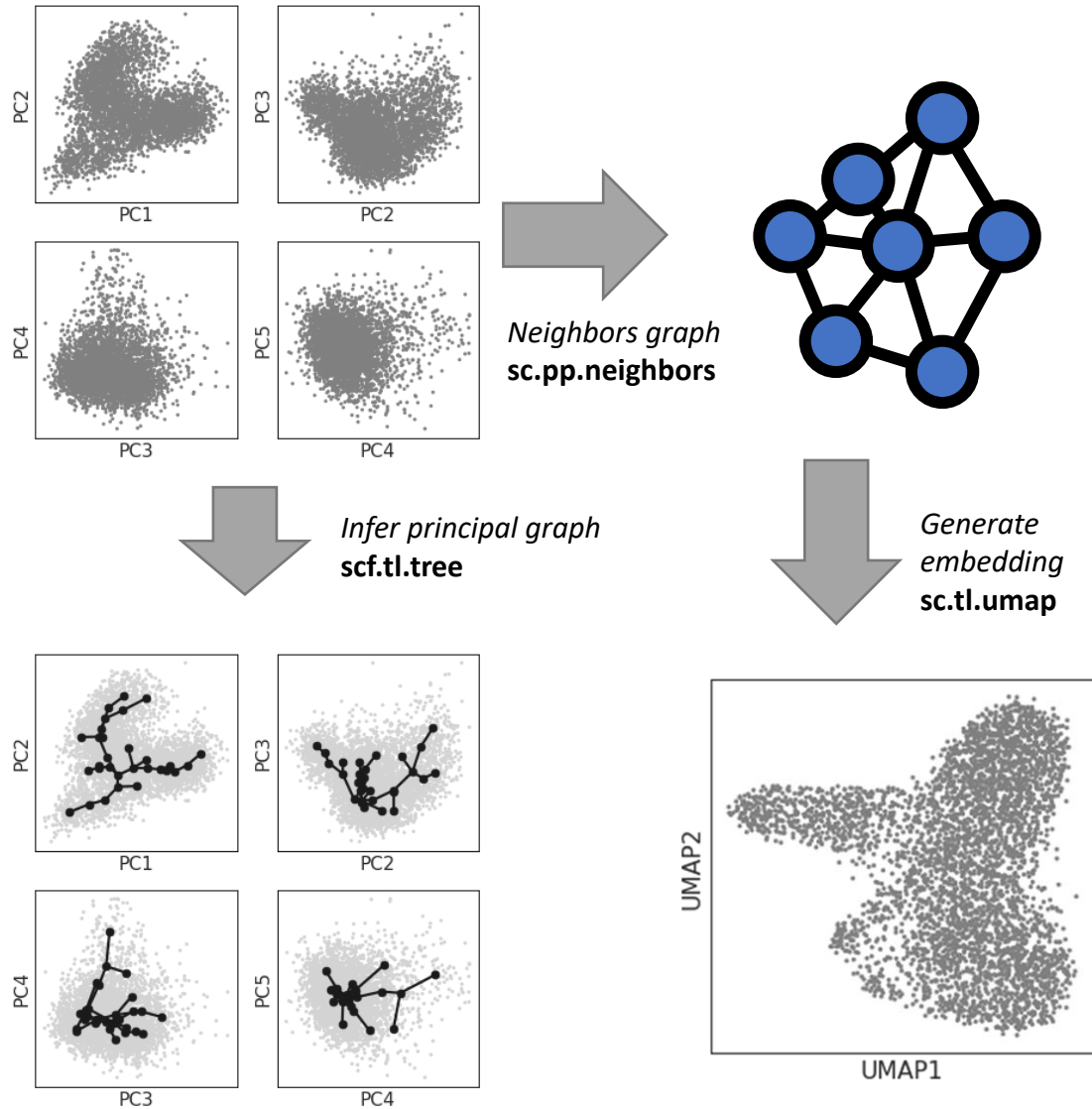


$\xleftarrow{\text{Random walk following a time dependent diffusion process, constrained by a gaussian kernel}}$

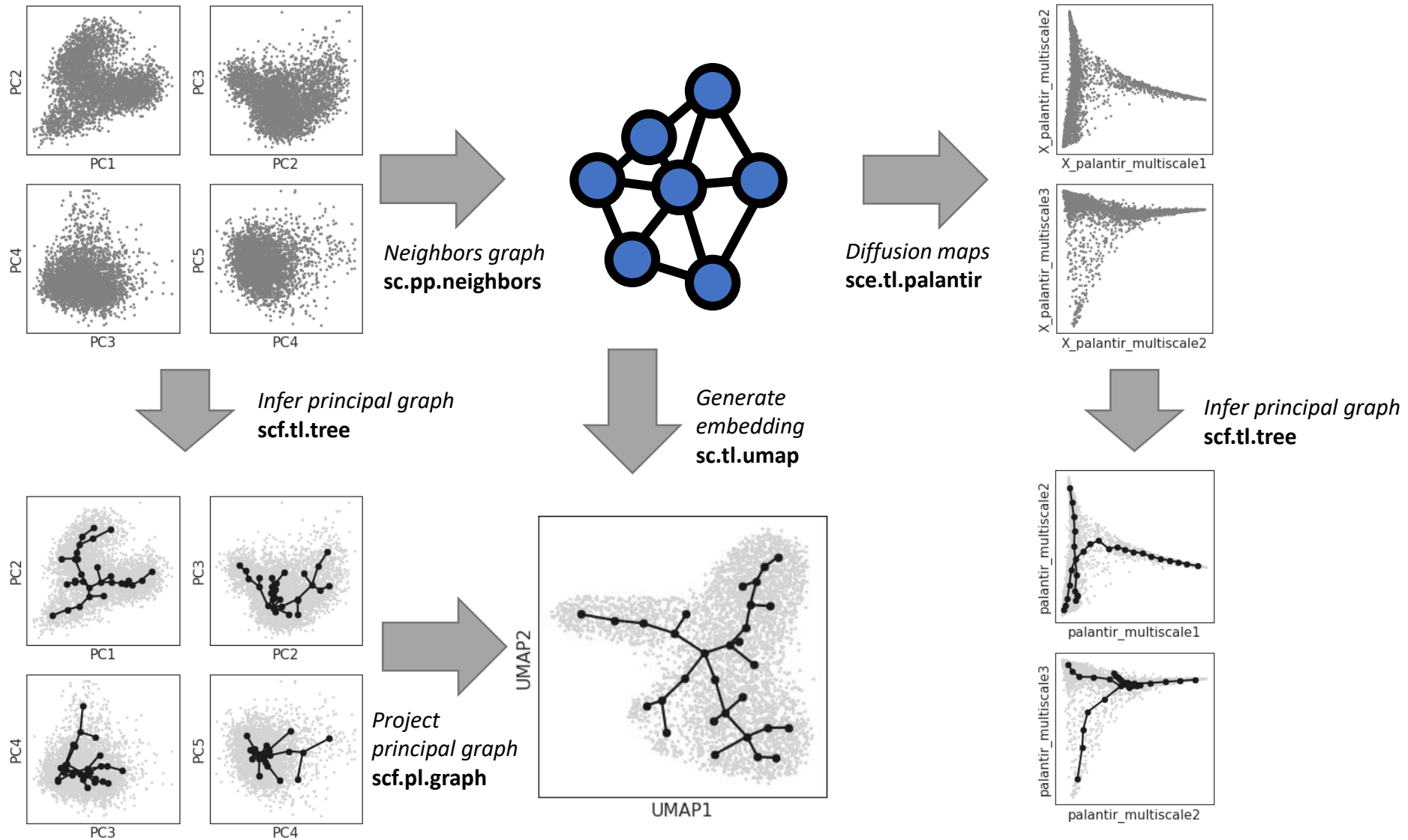
$n \times n$ Markovian transition probability matrix

Diffusion maps

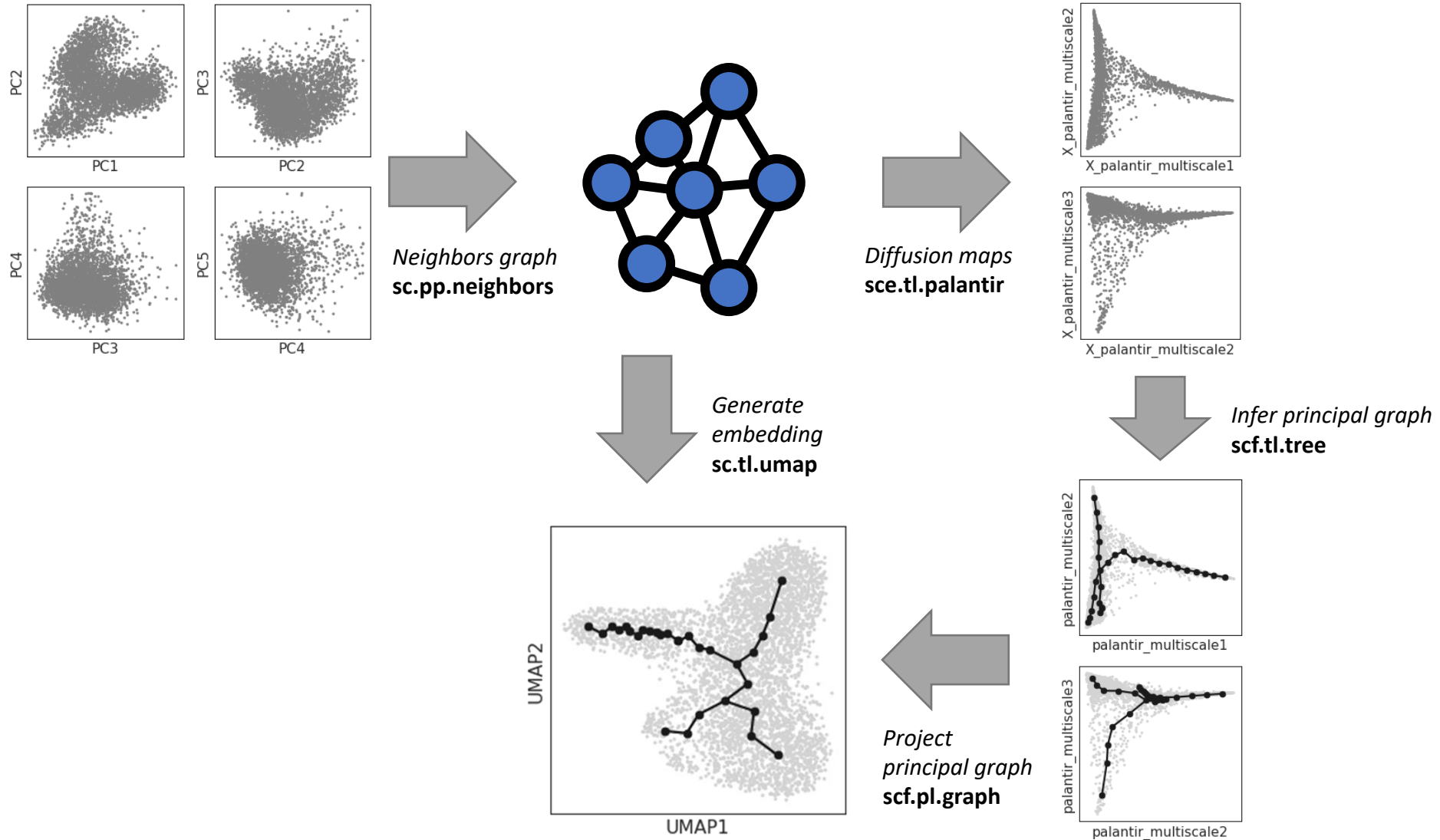
Trajectory inference in scRNAseq



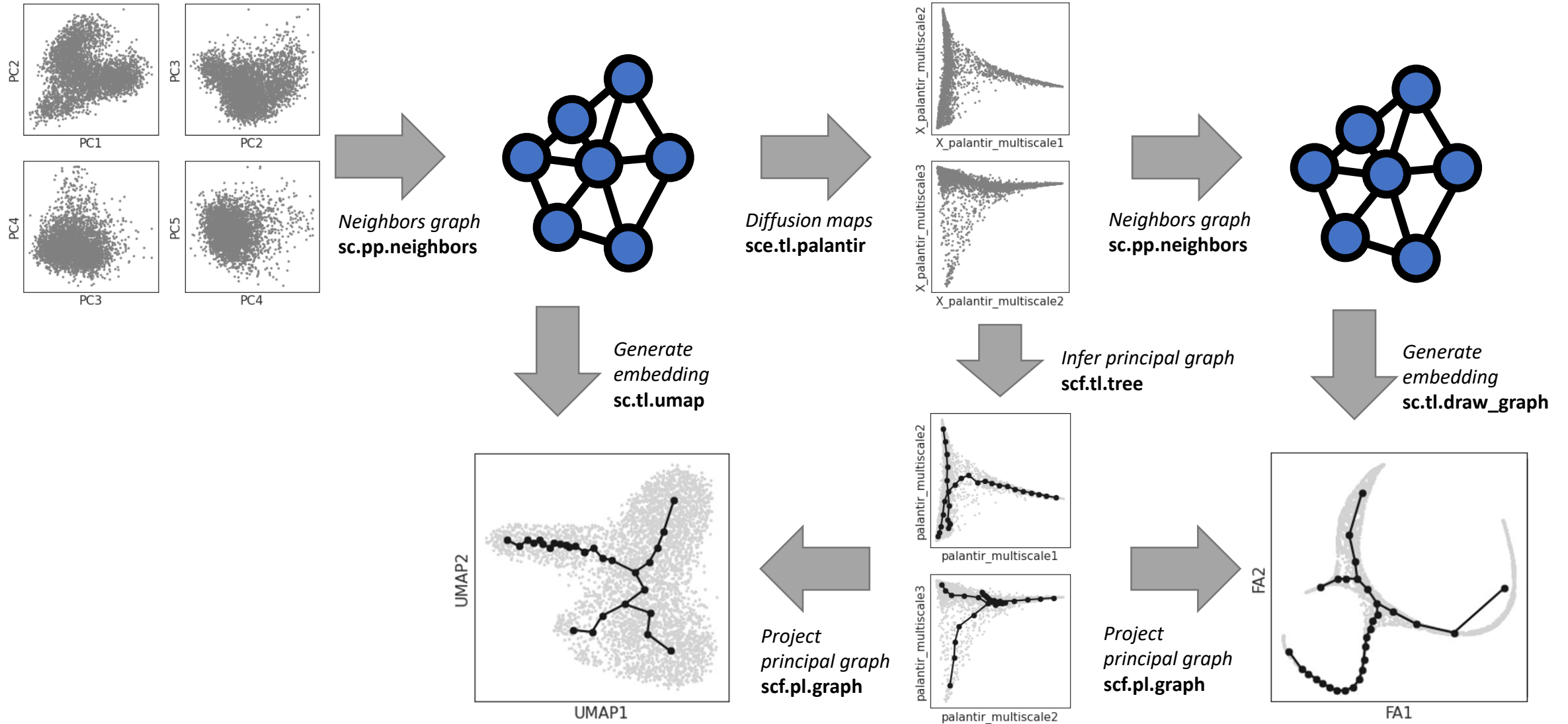
Trajectory inference in scRNAseq



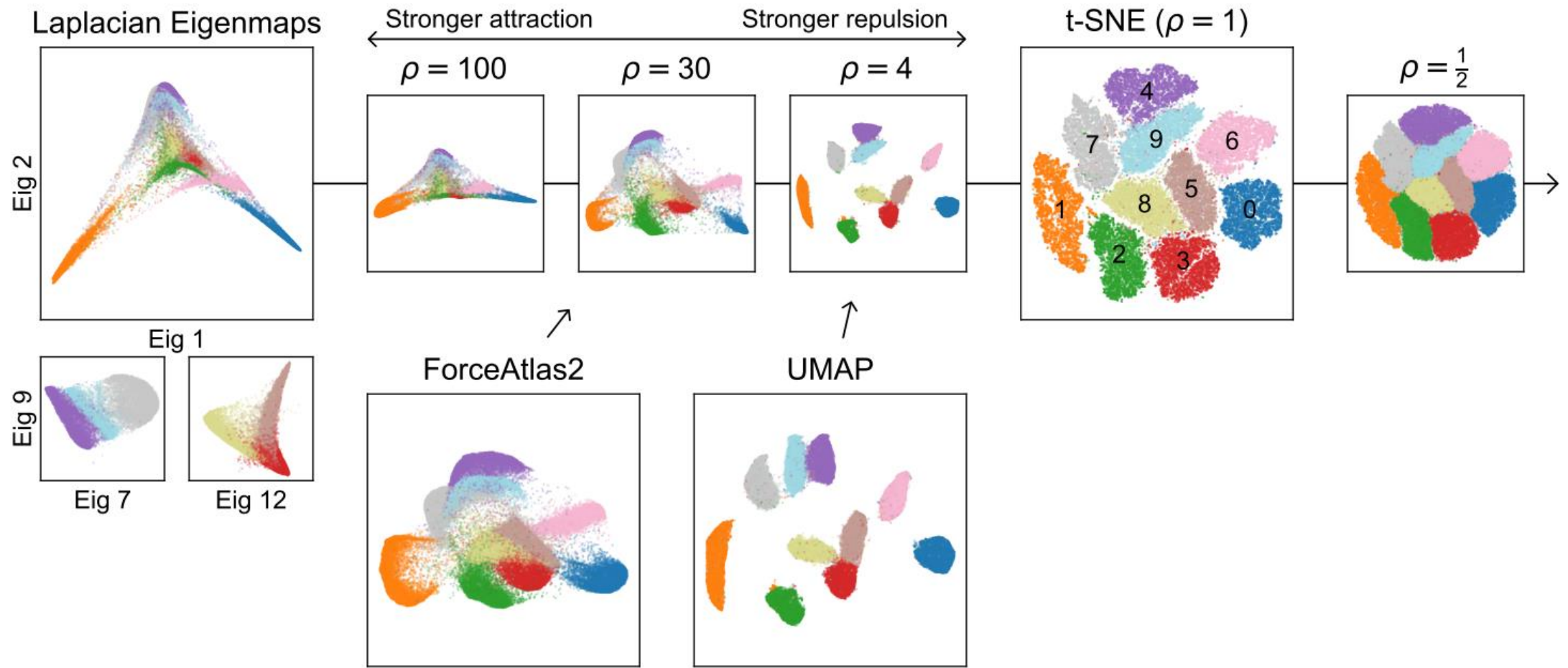
Trajectory inference in scRNAseq



Trajectory inference in scRNAseq



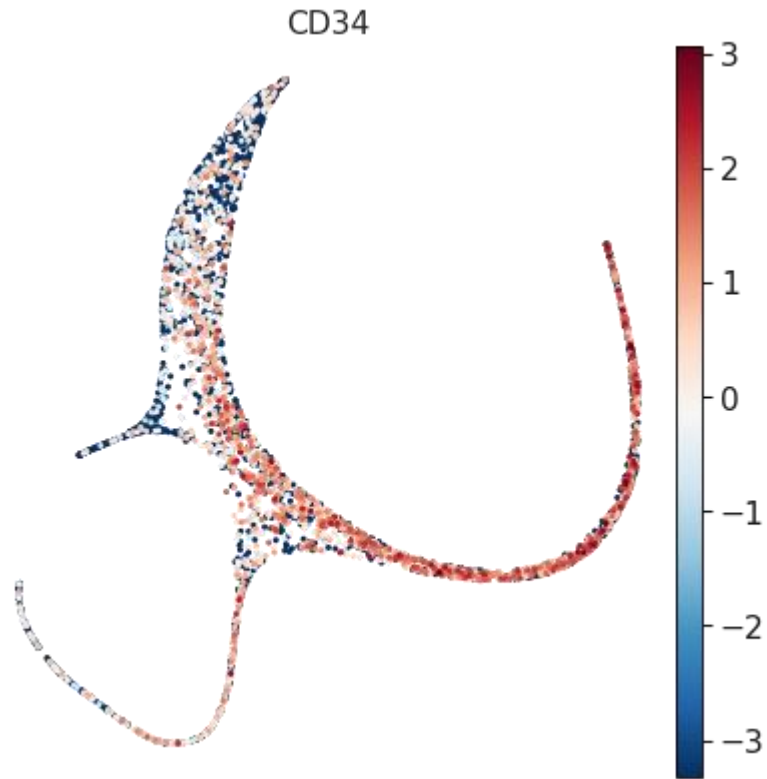
tSNE? UMAP? ForceAtlas2?



Must read!

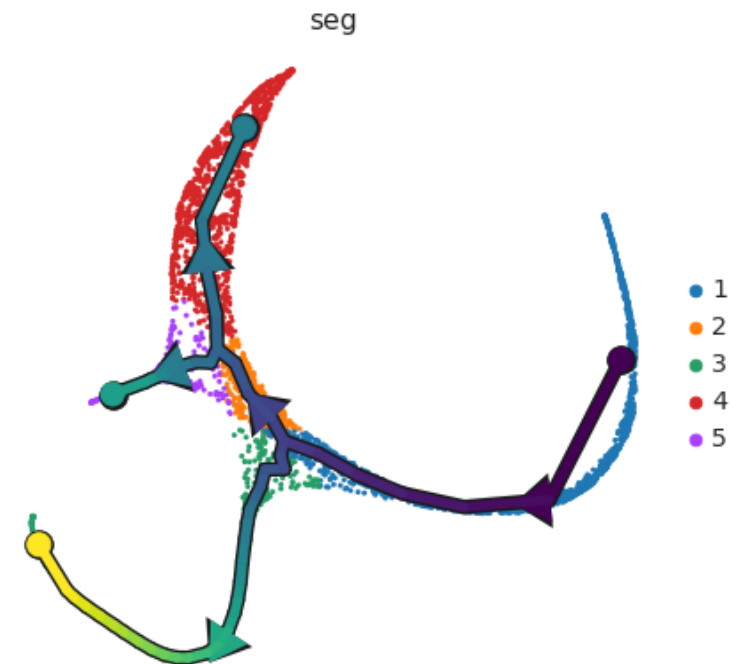
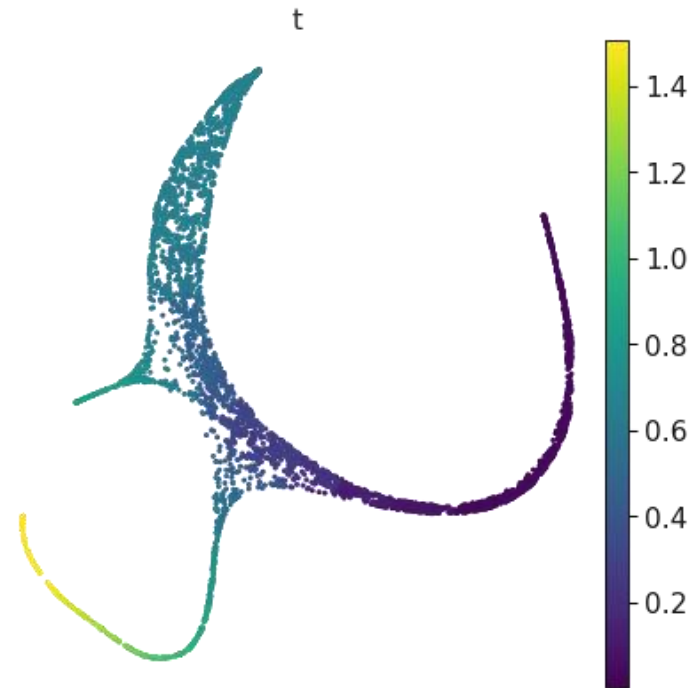
<https://arxiv.org/pdf/2007.08902.pdf>

Selecting the root



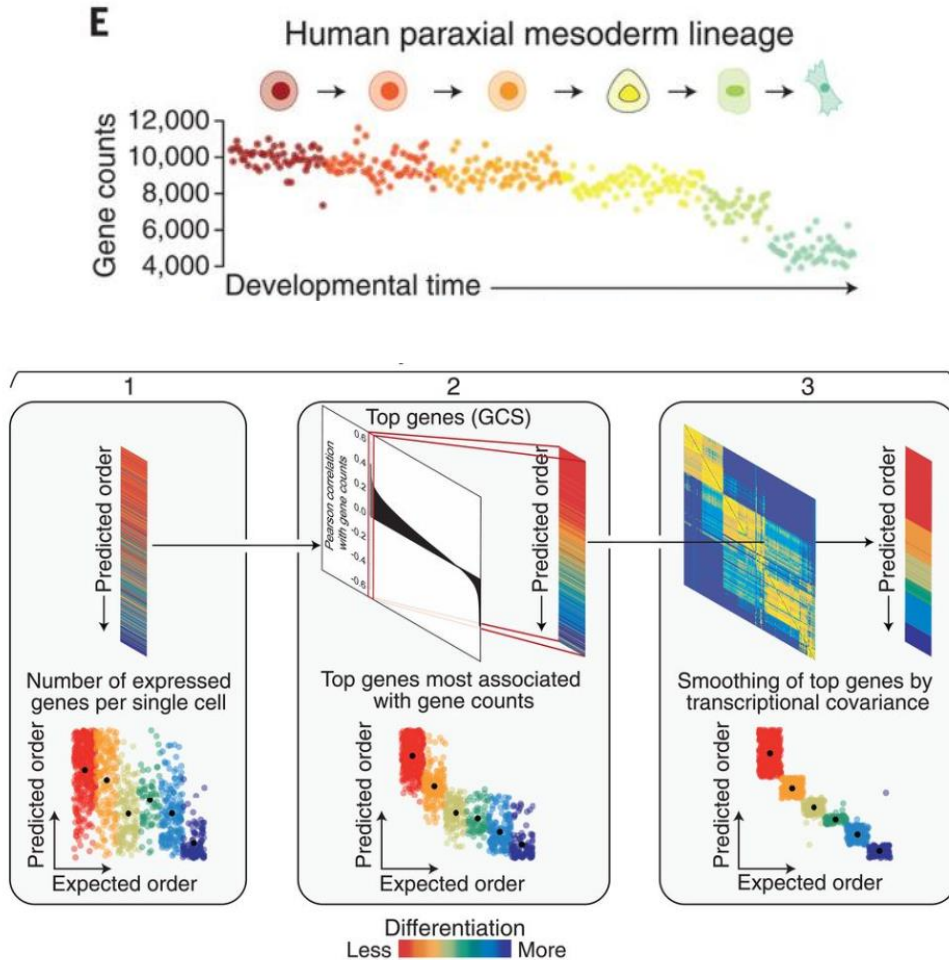
Use the help from:

- Experimental knowledge (sampling timepoint)
- Biological knowledge (known progenitor markers)



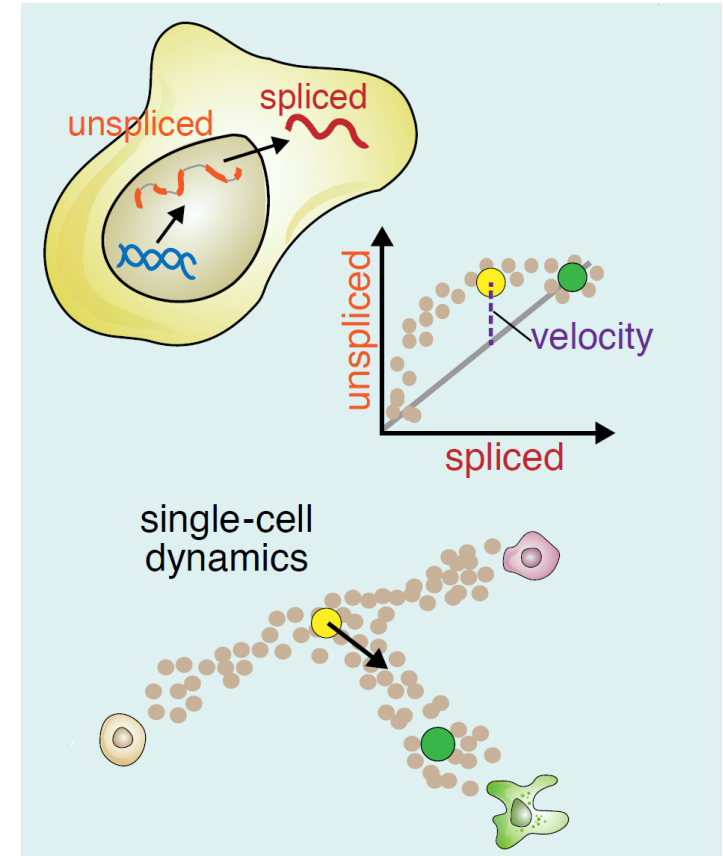
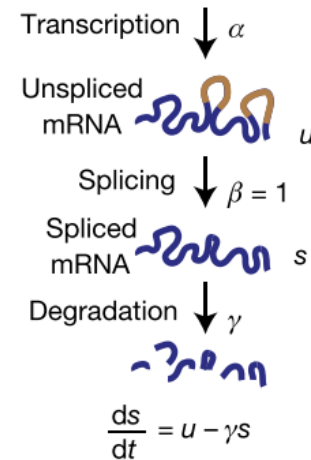
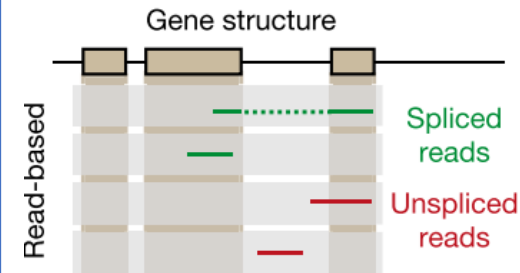
Need help finding the root?

CytoTRACE



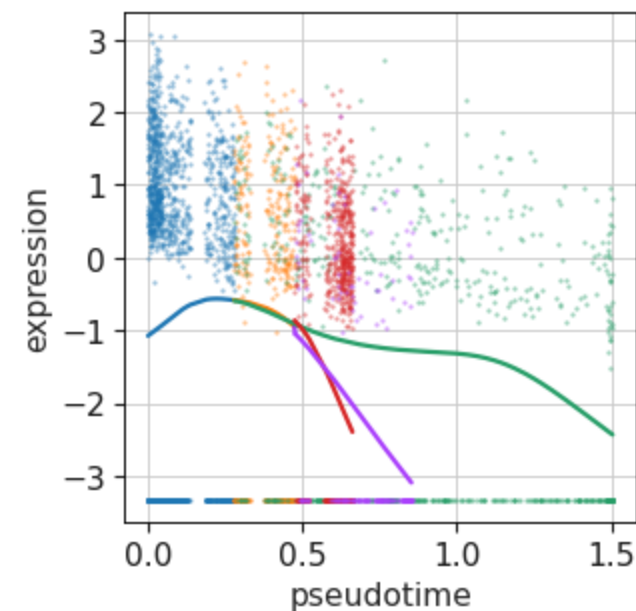
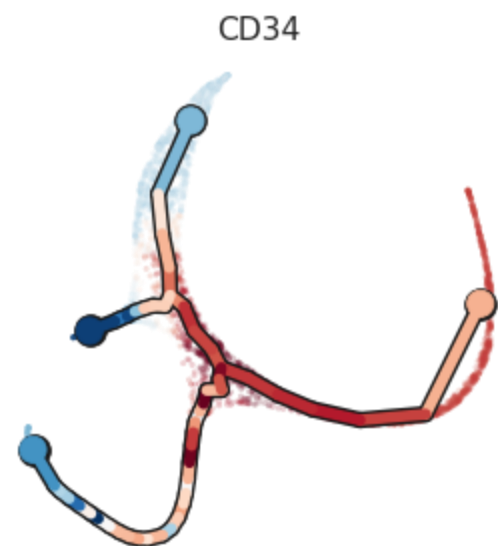
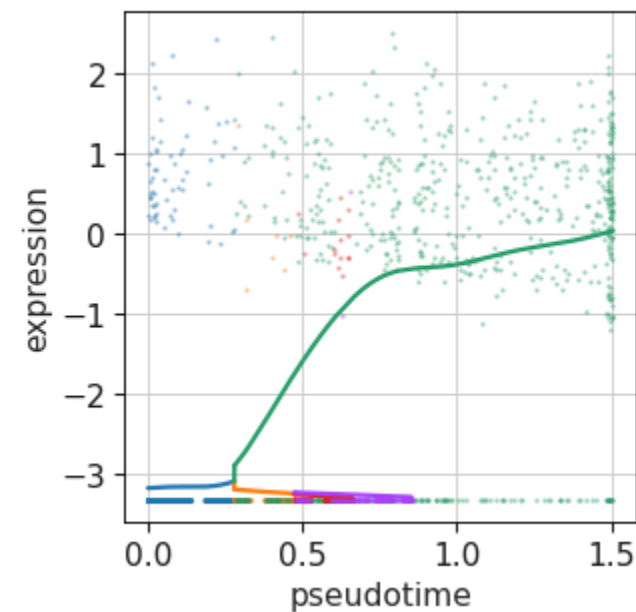
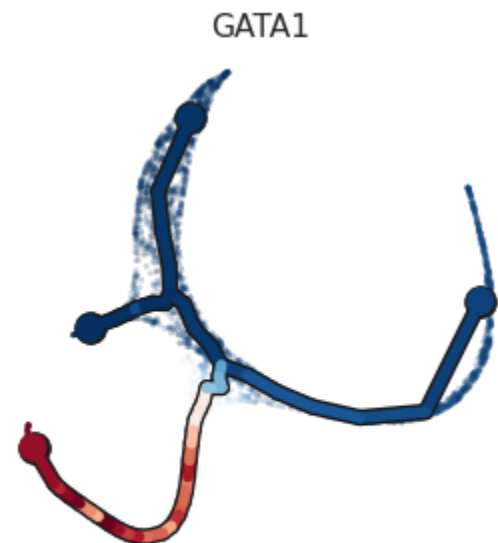
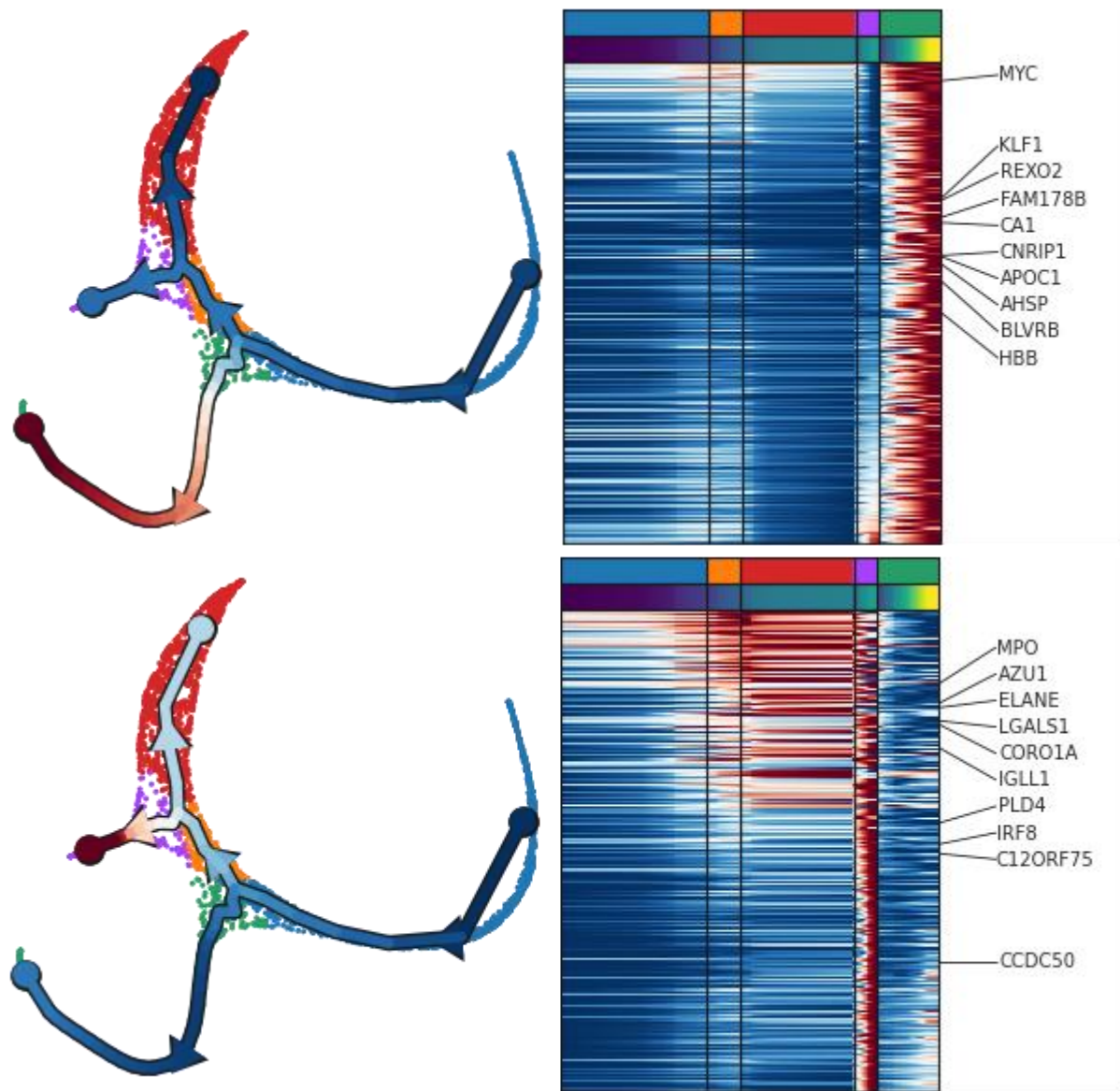
Gulati, G. S. *et al.* Single-cell transcriptional diversity is a hallmark of developmental potential. *Science* (80-.). (2020) doi:10.1126/science.aax0249.

RNA Velocity

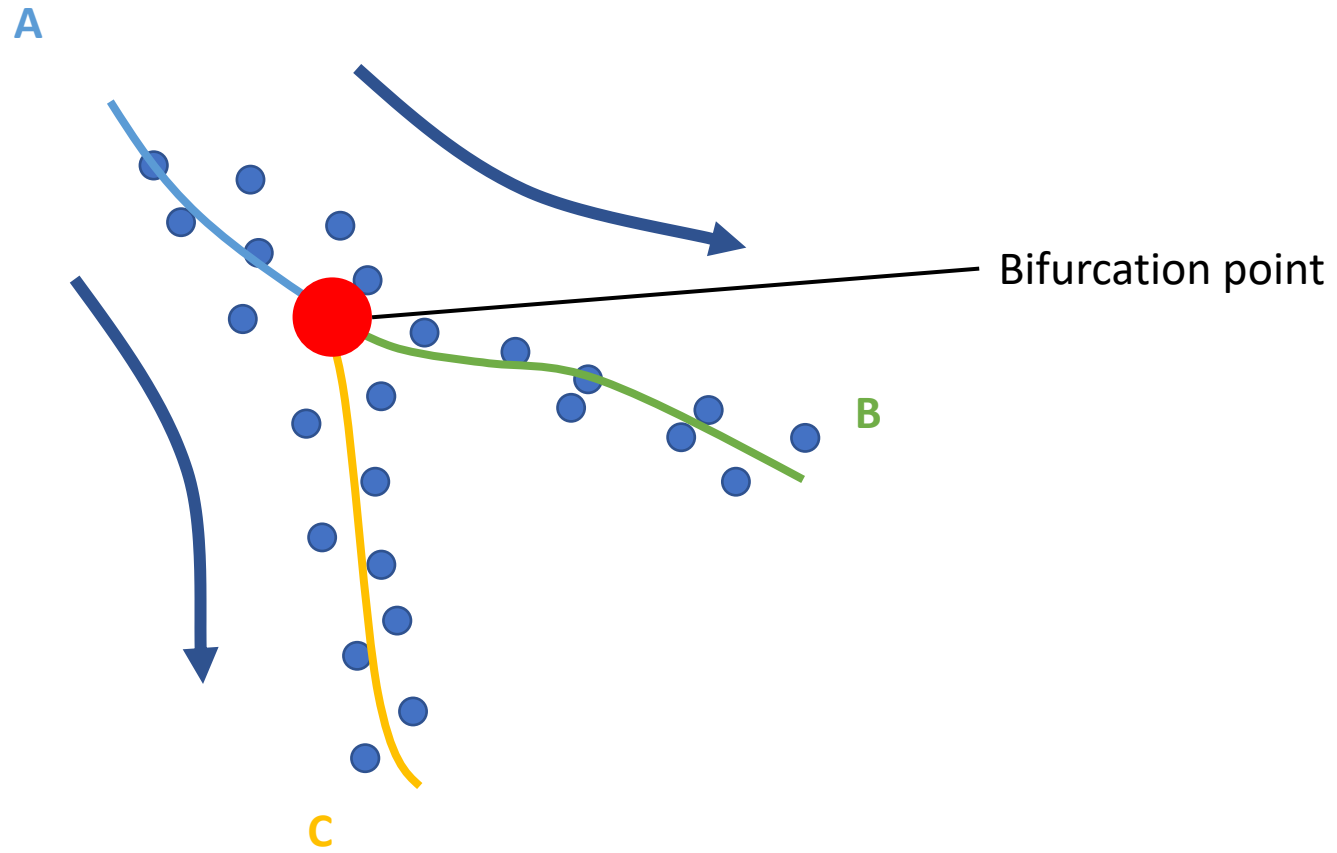


Edited from: Lederer, A. R. & La Manno, G. The emergence and promise of single-cell temporal-omics approaches. *Current Opinion in Biotechnology* (2020). doi:10.1016/j.copbio.2019.12.005

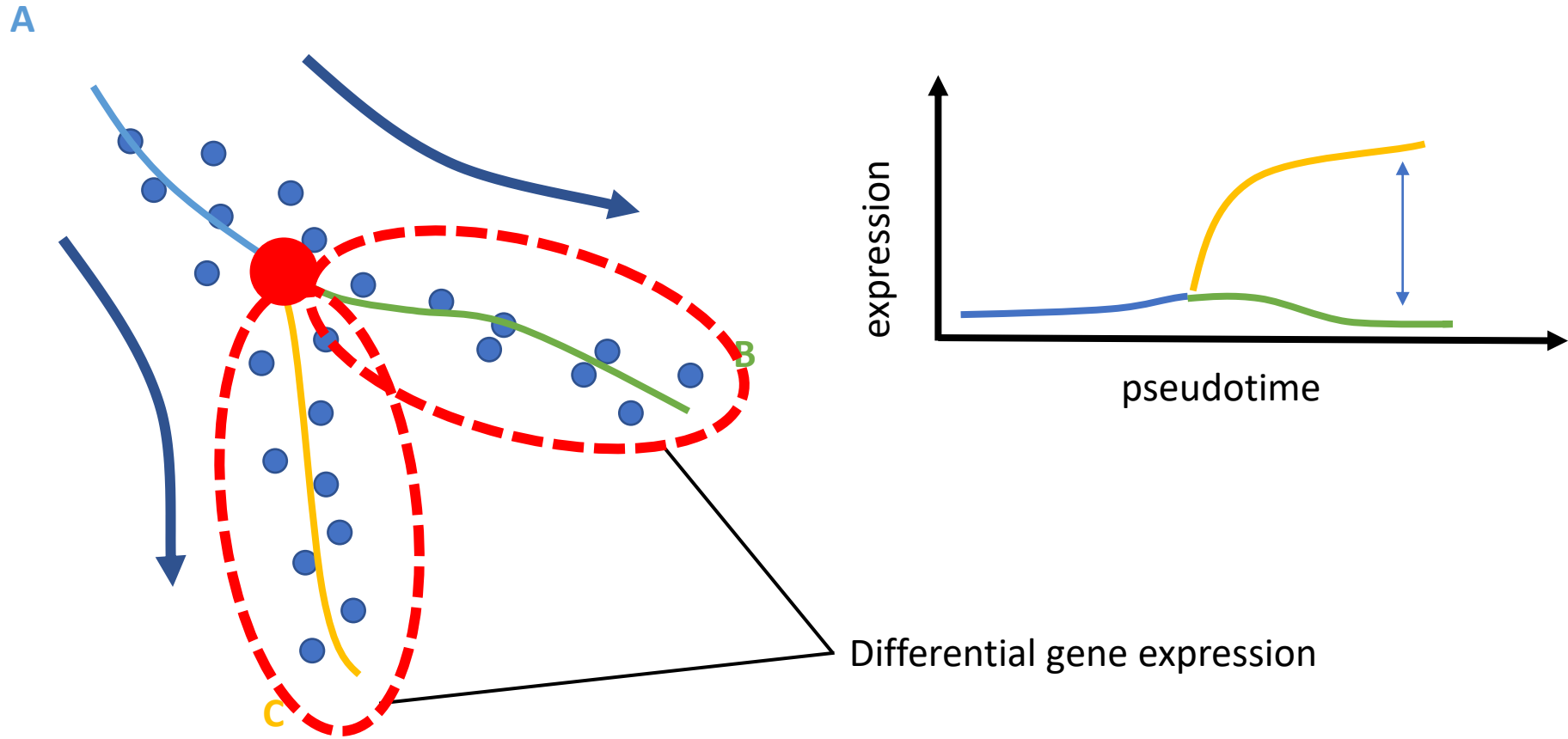
Recover the dynamics



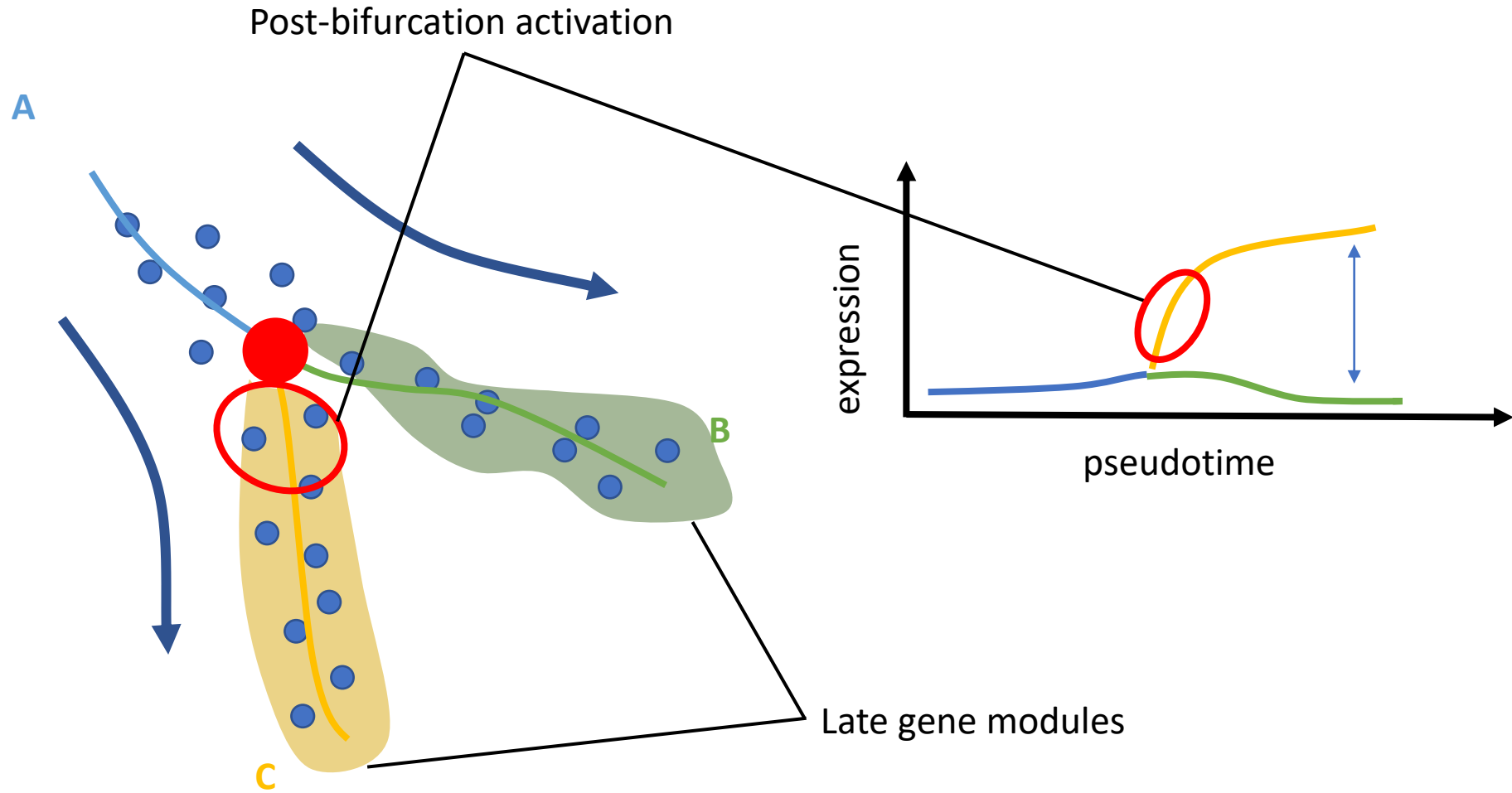
Probing bifurcations



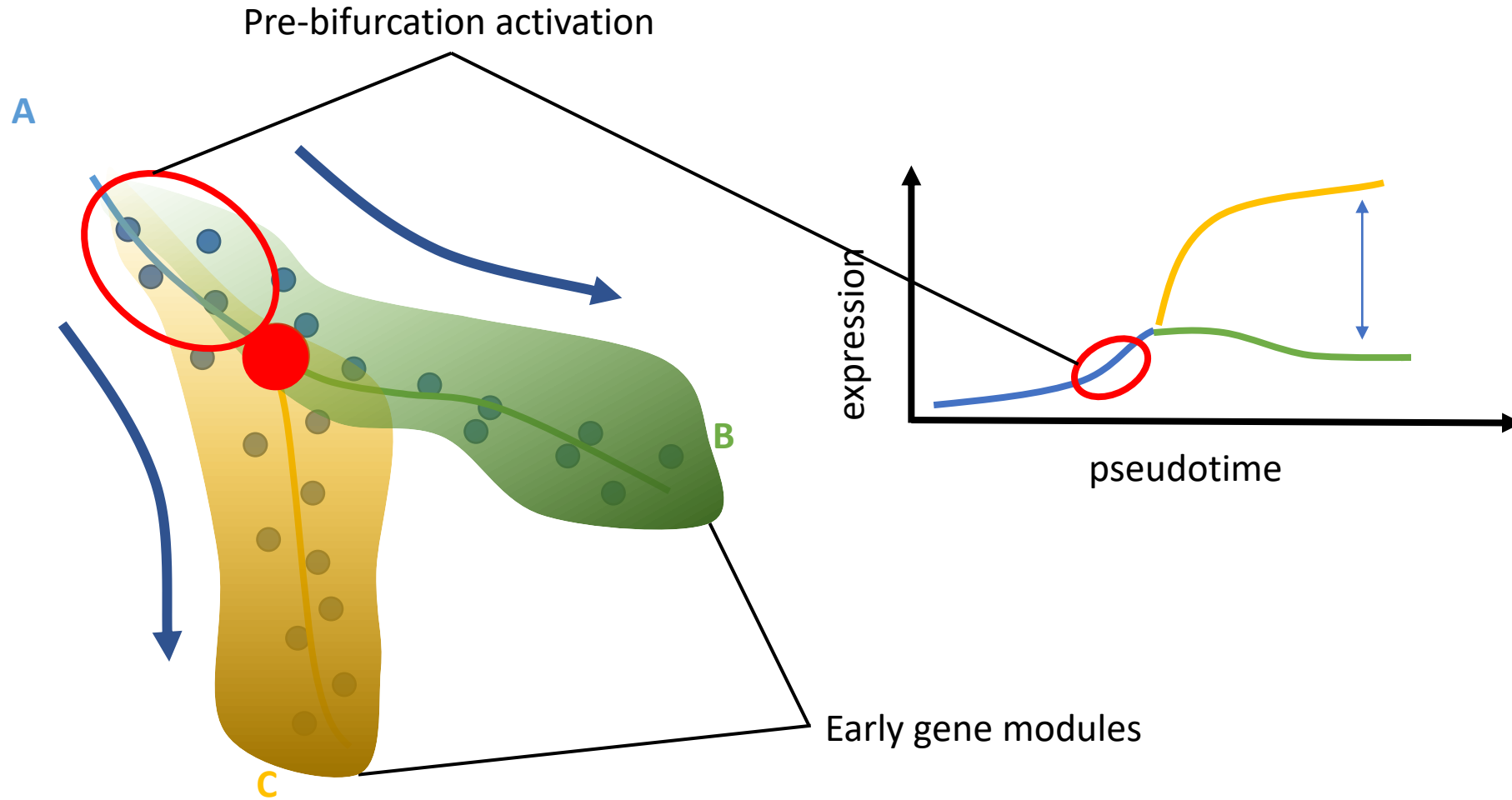
Probing bifurcations



Probing bifurcations

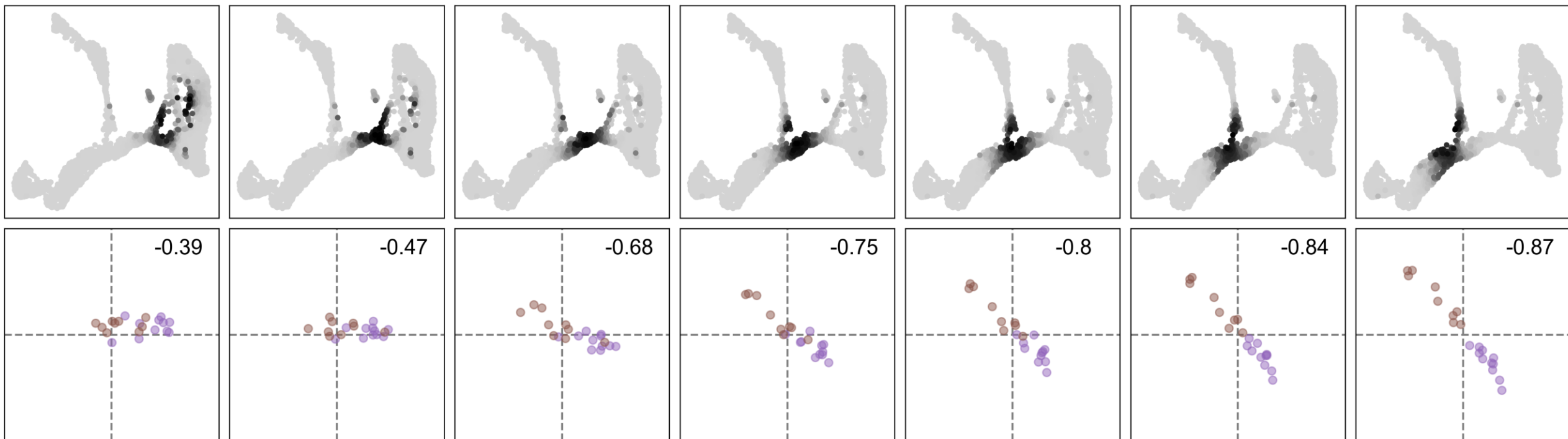
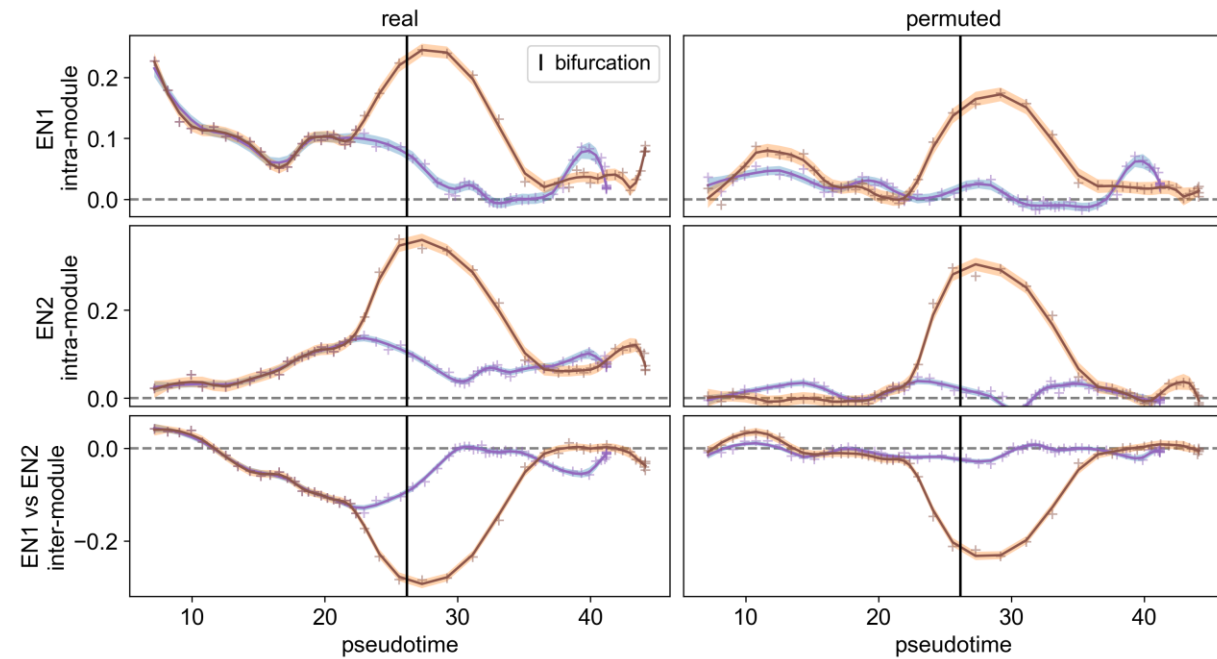


Probing bifurcations



Probing bifurcations

[https://scfates.readthedocs.io/en/latest/notebooks/Advanced bifurcation analysis.html#Bifurcation-analysis](https://scfates.readthedocs.io/en/latest/notebooks/Advanced%20bifurcation%20analysis.html#Bifurcation-analysis)



Thank you for listening!

scFates

Github repo: github.com/LouisFaure/scFates

Documentation: scfates.readthedocs.io

Package: pypi.org/project/scFates

Reproducibility: https://github.com/LouisFaure/Trajectory_Inference_workshop/

Soldatov *et al.* (2019), *Spatiotemporal structure of cell fate decisions in murine neural crest*, [Science](#).